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Synthesis of the Docosanasaccharide Arabinan Domain of Mycobacterial Arabinogalactan and a Proposed Octadecasaccharide Biosynthetic Precursor

Maju Joe, Yu Bai, Ruel C. Nacario, and Todd L. Lowary*

Contribution from the Alberta Ingenuity Centre for Carbohydrate Science and Department of Chemistry, The University of Alberta, Gunning-Lemieux Chemistry Centre, Edmonton, Alberta T6G 2G2, Canada

Received May 2, 2007; E-mail: tlowary@ualberta.ca

Abstract: Two major components of the cell wall in mycobacteria, including *Mycobacterium tuberculosis*, the causative agent of tuberculosis (TB), are polysaccharides containing arabinofuranose residues. In one of these polysaccharides, arabinogalactan, this arabinan domain consists of three identical motifs of 22 arabinofuranose residues, which are in turn attached to an underlying galactofuranan backbone. Recent studies have proposed that this docosanasaccharide motif, and a structurally related arabinan present in another cell wall polysaccharide, lipoarabinomannan, are biosynthesized from a common octadecasaccharide precursor. To facilitate the testing of this hypothesis, we report here the first total syntheses of these 18- and 22-residue oligosaccharides both functionalized with an aminooctyl linker arm. The route to the target compounds involved the preparation of four tri- to heptasaccharide building blocks possessing only benzoyl protecting groups that were coupled in a highly convergent manner via glycosyl trichloroacetimidate donors. Each of the targets could be prepared in only six steps from these intermediates, and in both cases more than 10 mg of material was obtained. These compounds are expected to be useful tools in probing the biosynthesis of these arabinan-containing polysaccharides. Such studies are essential prerequisites for the identification of novel anti-TB agents that target arabinan assembly.

Introduction

Mycobacterial infections have received significant attention recently due to their increasing incidence in industrialized countries as well as the emergence of drug-resistant strains of *Mycobacterium tuberculosis*, the organism that causes the most high profile of these diseases, tuberculosis (TB).¹ Notably, in the past year the identification of “extreme” drug-resistant TB (XDR-TB) has raised considerable alarm.² Successful treatment of TB requires a regimen of multiple antibiotics that must be administered over a number of months,³ and failure to complete this process is a major cause of drug resistance.⁴ Such protracted treatments are necessitated by the unusual structure of the mycobacterial cell wall, which promotes bacterial survival by acting both as a permeability barrier to the passage of antibiotics and also as a modulator of the host immune system.⁵ New drugs to treat TB and other mycobacterial diseases are therefore of

great current interest,⁶ and the enzymes involved in the biosynthesis of cell wall components are particularly attractive targets for antibiotic action. Indeed, two of the frontline antibiotics used to treat TB, ethambutol and isoniazid, act by preventing the formation of an intact cell wall.⁷

The major structural component of the cell wall complex is the mycolyl–arabinogalactan–peptidoglycan complex, which consists of an arabinogalactan (AG) polysaccharide attached, at its nonreducing end, to mycolic acids and, at its reducing end, to peptidoglycan.⁸ The linkage between the AG and peptidoglycan is through a disaccharide phosphate moiety (α -L-Rhap-(1 \rightarrow 4)- α -D-GlcpNAc-O-PO₃) to which is attached a chain of \sim 30 D-galactofuranose residues, polymerized via alternating β -(1 \rightarrow 5) and β -(1 \rightarrow 6) linkages. Along this galactan chain are three pendant arabinan chains,⁹ each containing 22 D-arabinofuranose (Araf) residues, in a combination of α -(1 \rightarrow 5), α -(1 \rightarrow 3), and β -(1 \rightarrow 2) linkages, to which the mycolic acids are esterified at the nonreducing end.¹⁰ In addition, recent work¹¹

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has confirmed an earlier report¹² that the arabinan domain in some mycobacterial species is further functionalized by the addition of substoichiometric amounts of a single galactosamine residue.

A second important cell wall molecule is lipoarabinomannan (LAM), a major antigen and immunomodulatory molecule.¹³ LAM consists of a phosphatidylinositol moiety linked to an α -(1 \rightarrow 6)-linked mannopyranan core to which is attached an arabinan similar in structure to that found in AG.¹³ LAM can be further elaborated with species-specific capping motifs (e.g., inositol phosphate residues,¹⁴ mannopyranosyl oligosaccharides,¹⁵ or 5-thiomethyl-D-xylofuranose residues¹⁶), which are attached at the nonreducing end of the arabinan. Succinate moieties are also found as modifications in LAM.¹⁷

Although both AG and LAM contain an arabinan domain of similar structure, differences between these motifs exist. In particular, increasing evidence has now accumulated to suggest that while the arabinan domain in AG is structurally well defined (i.e., a 22-residue motif), the one present in LAM is more heterogeneous.^{13,18} Despite significant advances in recent years, mycobacterial arabinan biosynthesis is still relatively poorly understood.^{8,13,19} The majority of the *Araf* residues appear to be installed by a family of arabinosyltransferases (AraTs), EmbA, EmbB, and EmbC,²⁰ which utilize decaprenolphospho-arabinose (DPA)²¹ as the donor species. It has been proposed that AG biosynthesis is mediated by EmbA and EmbB, while EmbC is involved in LAM biosynthesis.^{20b,c} Efforts to purify or recombinantly express these enzymes have not been successful, but it is possible to assay their activity in a mycobacterial membrane preparation.²² Enzymes other than the Emb proteins may also be involved, and indeed Besra and co-workers have reported that a distinct AraT, designated AftA, installs the first *Araf* residue attached to the galactan, thus “priming” the molecule for full-length arabinan assembly.²³ In addition, a very recent study²⁴ has demonstrated that another enzyme, termed AftB, is involved in arabinan biosynthesis, and the available

data point to its role as a β -(1 \rightarrow 2)-AraT that installs the terminal *Araf* residues in the arabinan.

As part of the increasing body of work on mycobacterial arabinan biosynthesis, Chatterjee and co-workers have proposed¹⁸ that a key intermediate in the biosynthesis of the arabinan domain of both AG and LAM is a motif (**1**, Figure 1) containing 18 *Araf* residues attached via α -(1 \rightarrow 5) and α -(1 \rightarrow 3) linkages. It was further proposed that the complete 22-residue arabinan motif present in AG (**2**) would result from the addition of four β -(1 \rightarrow 2)-linked *Araf* residues by EmbA/EmbB, whereas the action of EmbC on **1** would lead to LAM arabinan. In light of the recent identification of AftB,²⁴ this enzyme may also be involved in the production of a complete arabinan domain in one or both of these polysaccharides.

Testing these hypotheses would be facilitated by access to milligram quantities of authentic samples of these motifs, which can be most conveniently obtained by chemical synthesis. Thus, we report here synthesis of these motifs, functionalized at the reducing end with an octylamino linker, which can be used for their conjugation to a solid support or in the preparation of neoglycoconjugates. This study represents the first synthesis of these motifs, and the targets **3** and **4** (Figure 1) are some of the largest and most complex oligosaccharides produced by chemical synthesis to date.²⁵

Results and Discussion

Impressive syntheses of large fragments of mycobacterial LAM have recently been reported.^{25d,26,27} The Seeberger group has synthesized a dodecasaccharide fragment containing six α -*Araf* and six α -mannopyranose (α -Manp) residues via a [6 + 6] strategy in which a key step is the coupling of the mannan and arabinan domains.²⁶ An even larger (28-residue) fragment, possessing the inositol, 15 α -Manp, and 12 α -*Araf* residues was synthesized via a route in which the key glycosylation was a [12 + 16] coupling between an arabinomannan donor and a mannosylated inositol acceptor.^{25d} These are elegant approaches to complicated molecules; however, neither contain the β -*Araf* residues present in the arabinan domain of mycobacterial AG. Introduction of these β -*Araf* residues in a stereocontrolled manner is challenging²⁸ and was a key consideration in planning our approach to the targets **3** and **4**.

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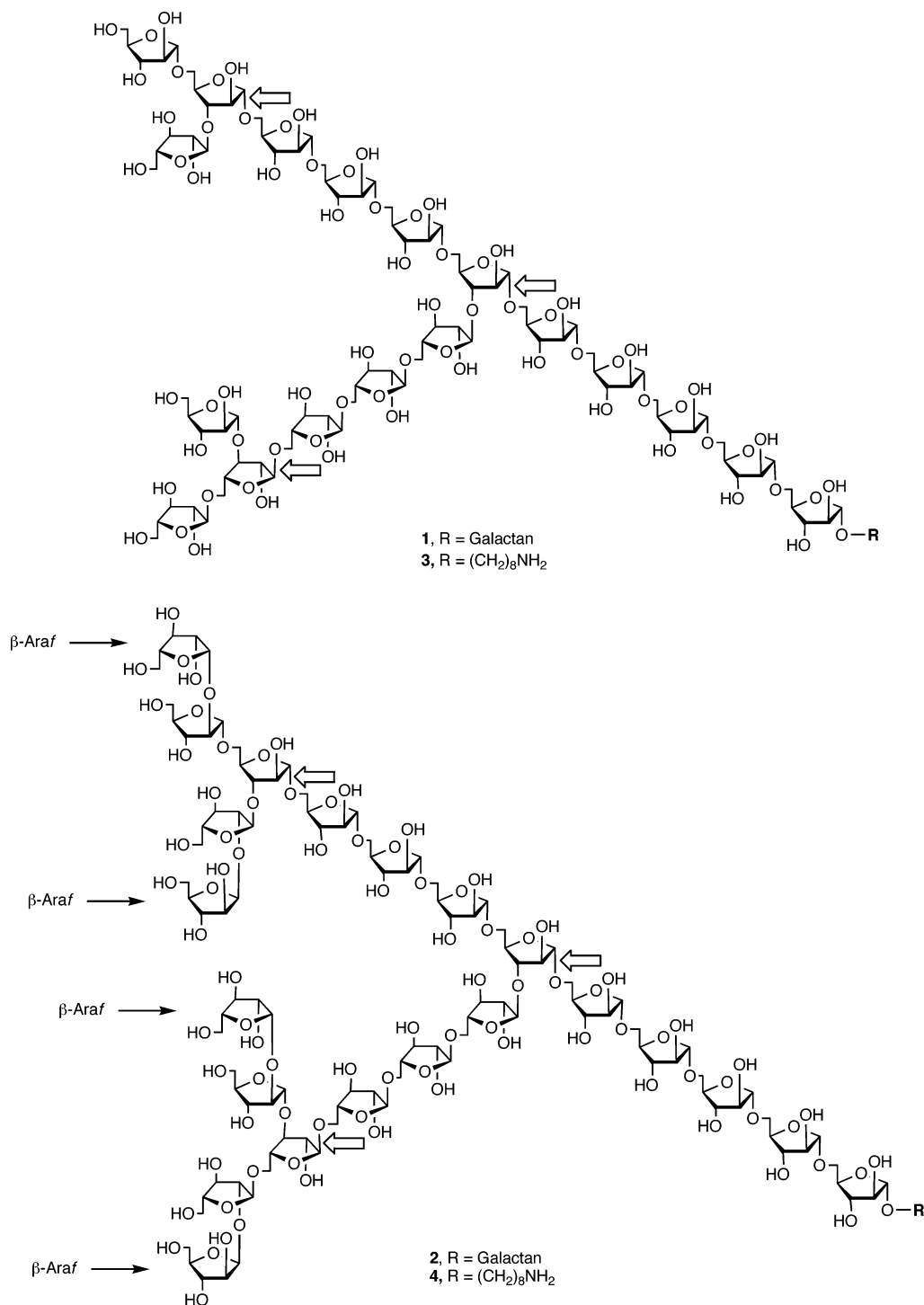


Figure 1. Araf₂₂ domain of mycobacterial AG (2) and the proposed Araf₁₈ biosynthetic precursor (1) and amino-octyl glycoside analogues of these motifs (3 and 4). The four β -Araf residues in 2 and 4 have been identified for clarity, and key synthetic disconnections are identified by open arrows.

In designing a route to 3 and 4, we determined that a convergent approach, in which the molecules would be assembled via the sequential coupling of tri- to heptasaccharide building blocks, would be the most efficient. This would allow us to prepare large amounts of these building blocks that, due to their relatively small size, could be readily characterized by NMR spectroscopy. Furthermore, by adding blocks of residues in the glycosylations, we expected that the resulting large changes in molecular weight would facilitate chromatographic separations of reaction mixtures. With regard to the installation of the four β -Araf residues in 4, we chose to prepare a building

block with these linkages already in place, as we envisioned that the separation of any unwanted isomers on relatively small intermediates would be easier than on larger structures. Finally, given the size of the final target molecules, we anticipated that the removal of a large number of benzyl groups at the end of the synthesis could be problematic, and therefore, a key part of the strategy was to use building blocks protected only with acyl (benzoyl) groups.

With these considerations in mind, key disconnections were identified for each molecule (open arrows in Figure 1) and four key building blocks were chosen (5–8, Figure 2). We envisioned

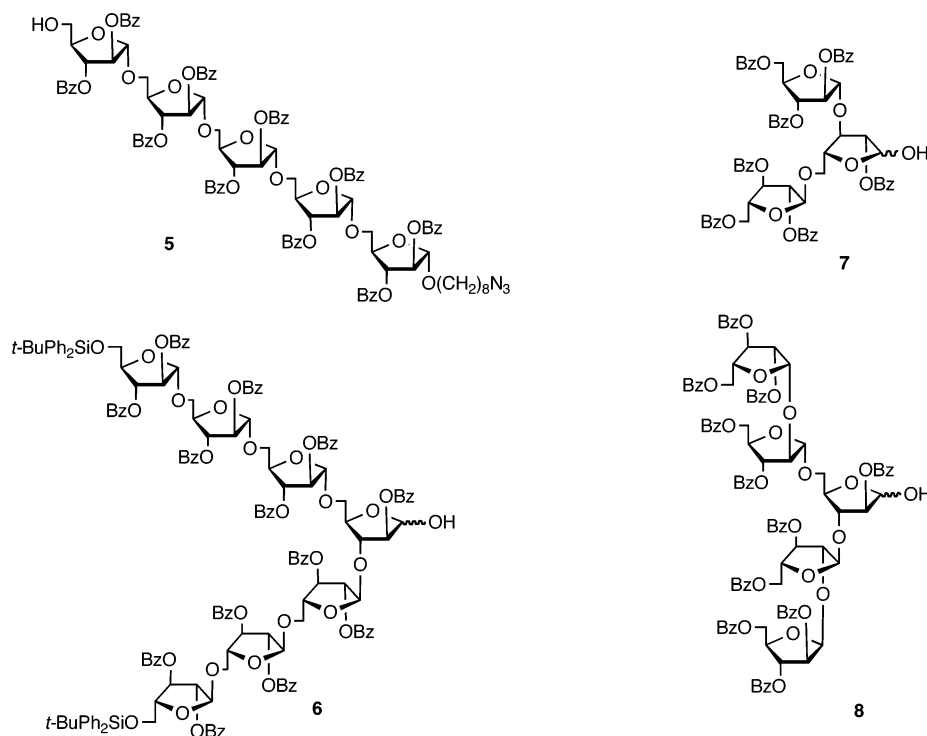


Figure 2. Key building blocks 5–8 required for the synthesis of 3 and 4.

that **3** could be obtained by glycosylation of pentasaccharide **5** with a trichloroacetimidate derived from heptasaccharide **6** to give a dodecasaccharide. Cleavage of the silyl ethers in this Araf₁₂ species would provide a diol that could be reacted with an excess of the trichloroacetimidate donor prepared from trisaccharide **7**, in turn yielding a fully benzoylated octadecasaccharide. Treatment with base followed by reduction would afford **3**. A similar approach could be used to access **4** by the use, in the final glycosylation reaction, of a trichloroacetimidate obtained from pentasaccharide **8**, in which the key β -Araf residues are already in place. In the coupling of all these building blocks, the stereochemistry of the newly formed glycosidic linkage would be controlled by neighboring group participation of the benzoyl group at C-2 of the donor.

Synthesis of Pentasaccharide 5. Having developed a general strategy to access the targets, we first addressed the synthesis of the building block that would become the reducing end of both **3** and **4**: pentasaccharide **5** (Scheme 1). The synthesis of **5** started from the protected thioglycoside **9**, which was prepared from D-arabinose in six steps as previously reported.²⁹ Conversion of **9** into glycoside **10** was carried out by reaction with 8-azido-1-octanol³⁰ and *N*-iodosuccinimide–silver triflate (NIS–AgOTf).³¹ The product was obtained in 92% yield, and the anomeric stereochemistry was determined, as was the case for all glycosylations reported here, using NMR spectroscopy.³² Consistent with the assigned α -stereochemistry, in the ¹H NMR spectrum for **10** the signal for H-1 appeared as a singlet at 5.24 ppm and in the ¹³C NMR spectrum the C-1 resonance was at

105.6 ppm. Deprotection of **10** to give **11** was achieved upon reaction with HF–pyridine, a transformation that proceeded in 83% yield. Thioglycoside **9** was also hydrolyzed by reaction with NIS–AgOTf in aqueous THF, which afforded lactol **12** in 85% yield as a 1:1 α/β mixture.

With a route to **12** in place, it was reacted with DBU and trichloroacetonitrile, yielding the trichloroacetimidate **13**, which, following a brief purification step, was used to glycosylate the known thioglycoside alcohol **14**³³ using TMSOTf as the promoter.³⁴ The product disaccharide thioglycoside **15** was obtained in 76% yield over two steps. This glycosyl donor was then reacted with alcohol **11** using NIS–AgOTf promotion, which afforded an 84% yield of the fully protected trisaccharide **16**. Cleavage of the silyl ether protecting group under standard conditions (HF–pyridine) gave trisaccharide alcohol **17** (95% yield), which was subsequently reacted with an excess of disaccharide thioglycoside **15** to afford pentasaccharide **18**. This [2 + 3] glycosylation provided **18** in 86% yield. Conversion of **18** into building block **5** was achieved in 96% yield by treatment with HF–pyridine. In the ¹³C NMR spectrum of **5**, five anomeric carbons were observed between 105.5 and 105.8 ppm, thus establishing their α -stereochemistry.³²

Synthesis of Heptasaccharide 6. The precursor to the branched core of both **3** and **4** was heptasaccharide **6**, which was synthesized as shown in Scheme 2. First, thioglycoside **19**, which can be easily obtained in multigram quantities in three steps from D-arabinose,³⁵ was transformed in 90% yield to the corresponding *p*-methoxyphenyl (PMP) glycoside **20** under standard glycosylation conditions. Removal of the benzoyl groups in **20** was achieved by stirring in sodium methoxide and methanol, affording triol **21** in 87% yield. This triol was then

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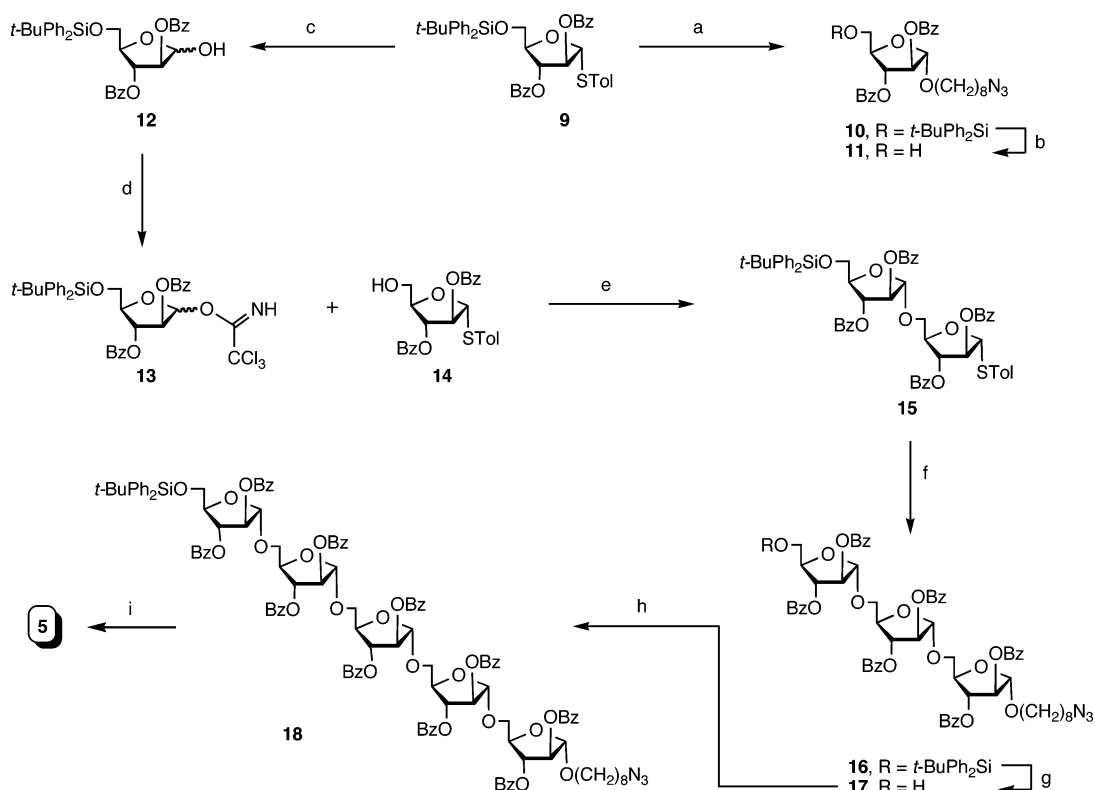
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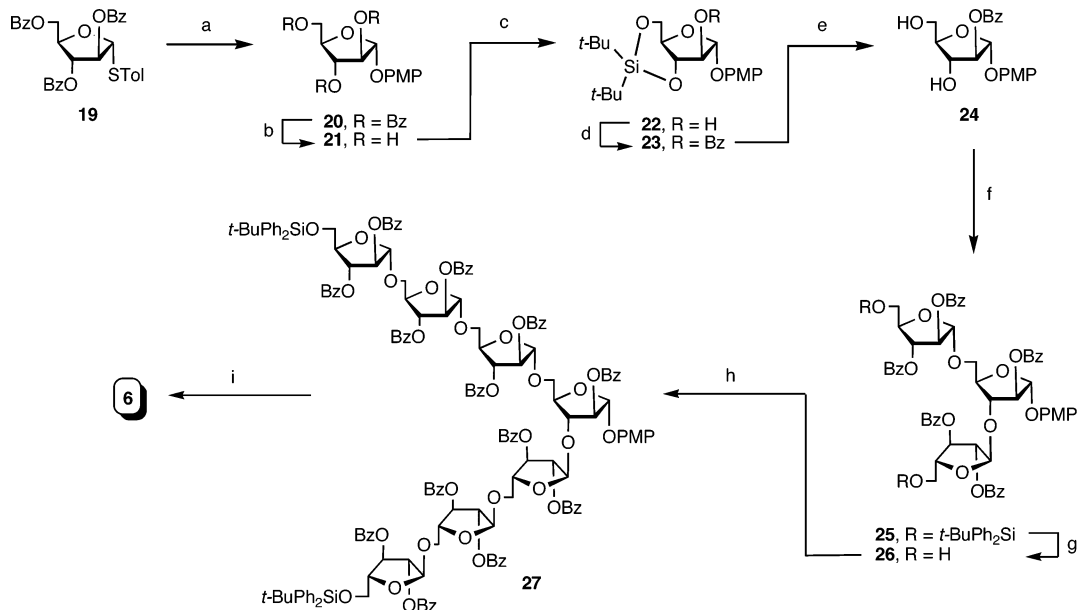
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Scheme 1^a

^a Reagents and conditions: (a) HO(CH₂)₈N₃, NIS, AgOTf, 0 °C, 92%; (b) HF–pyridine, THF, 0 °C → rt, 83%; (c) NIS, AgOTf, THF–H₂O, 0 °C, 85%; (d) CCl₃CN, DBU, CH₂Cl₂, 0 °C → rt; (e) TMSOTf, –30 to –5 °C, 76% over two steps; (f) 11, NIS, AgOTf, 0 °C, 84%; (g) HF–pyridine, THF, 0 °C → rt, 95%; (h) 15, NIS, AgOTf, 0 °C, 86%; (i) HF–pyridine, THF, 0 °C → rt, 96%.

Scheme 2^a

^a Reagents and conditions: (a) *p*-methoxyphenol, NIS, AgOTf, CH₂Cl₂, 0 → 10 °C, 90%; (b) NaOCH₃, CH₃OH–CH₂Cl₂, rt, 87%; (c) (*t*-Bu)₂Si(OTf)₂, 2,6-lutidine, CH₂Cl₂–DMF, 0 °C; (d) BzCl, pyridine, 0 °C → rt; (e) HF–pyridine, THF, 0 °C → rt, 76% over three steps; (f) 9, NIS, AgOTf, CH₂Cl₂, 0 °C, 74%; (g) HF–pyridine, THF, 0 °C → rt, 89%; (h) 15, NIS, AgOTf, CH₂Cl₂, 0 °C, 89%; (i) CAN, THF–H₂O, 0 °C, 80%.

converted to silyl acetal **22** upon reaction with di-*tert*-butylsilyl bis(trifluoromethanesulfonate) and 2,6-lutidine in a mixture of DMF and dichloromethane. An 87% yield of **22** was obtained. Benzoylation of **22** under conventional conditions provided the fully protected glycoside **23**, which was then partially depro-

tected upon treatment with HF–pyridine to give the key intermediate diol **24** in 76% yield over three steps from **21**.

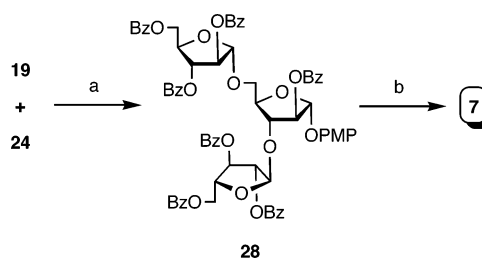
Glycosylation of **24** with an excess of thioglycoside **9** (see Scheme 1) afforded a 74% yield of the branched trisaccharide **25**, which was in turn deprotected (HF–pyridine), giving diol

26 in 89% yield. Elongation of this trisaccharide to the heptasaccharide was achieved by reaction of **26** with an excess of disaccharide thioglycoside **15** (Scheme 1) promoted by NIS–AgOTf. This reaction proceeded in 89% yield to give the expected fully protected heptasaccharide **27**, the ^{13}C NMR spectrum of which showed seven α -Araf anomeric carbons between 105.3 and 106.1 ppm. The final step in the preparation of **6** was the removal of the PMP group, which was achieved in 80% yield upon reaction with ceric ammonium nitrate (CAN) in aqueous THF.

Synthesis of Trisaccharide 7. The trisaccharide moiety at the nonreducing end of **3** was to be introduced via hemiacetal **7**, which was straightforwardly obtained as illustrated in Scheme 3. NIS–AgOTf-promoted glycosylation of diol **24** (Scheme 2) with an excess of thioglycoside **19** gave PMP trisaccharide **28** in 92% yield. Subsequent treatment with CAN in aqueous THF led to the formation of **7** as a 1:1 mixture of anomers in 85% yield.

Synthesis of Pentasaccharide 8. For the synthesis of the Araf₂₂ target **4**, the protected pentasaccharide **8** was needed for the motif at the nonreducing end of the molecule. A key issue in the preparation of this fragment was the installation of the two β -Araf moieties, which are stereochemically analogous to β -mannopyranosides.³⁶ A number of methods for the stereoselective formation of β -Araf linkages have been developed over the past several years by our group³⁷ and others.³⁸ For the preparation of **8**, we decided to employ the elegant method recently reported first by Boons and co-workers³⁹ and later by the Ito⁴⁰ and Crich⁴¹ groups, which relies on a conformationally rigidified thioglycoside donor.

The synthesis of **8** is shown in Scheme 4 and starts with thioglycoside **29**, which was prepared as previously described.⁴² Protection of the alcohol functionality in **29** as the levulinate ester (levulinic acid, DCC, DMAP in dichloromethane) afforded **30** in 90% yield. An excess of this fully protected thioglycoside was then coupled with the PMP glycoside diol **24** (Scheme 2) under our standard thioglycoside coupling conditions (NIS–AgOTf) to yield trisaccharide **31** in 76% yield. As expected, all three glycosidic linkages had the α -stereochemistry, as determined by ^{13}C NMR spectroscopy (three anomeric carbon signals at 106.2, 106.0, and 105.2 ppm). The resonances arising from the anomeric hydrogens of the residues bearing the silyl

Scheme 3^a

^a Reagents and conditions: (a) NIS, AgOTf, CH_2Cl_2 , 0 °C, 92%; (b) CAN, $\text{THF-H}_2\text{O}$, 0 °C, 85%.

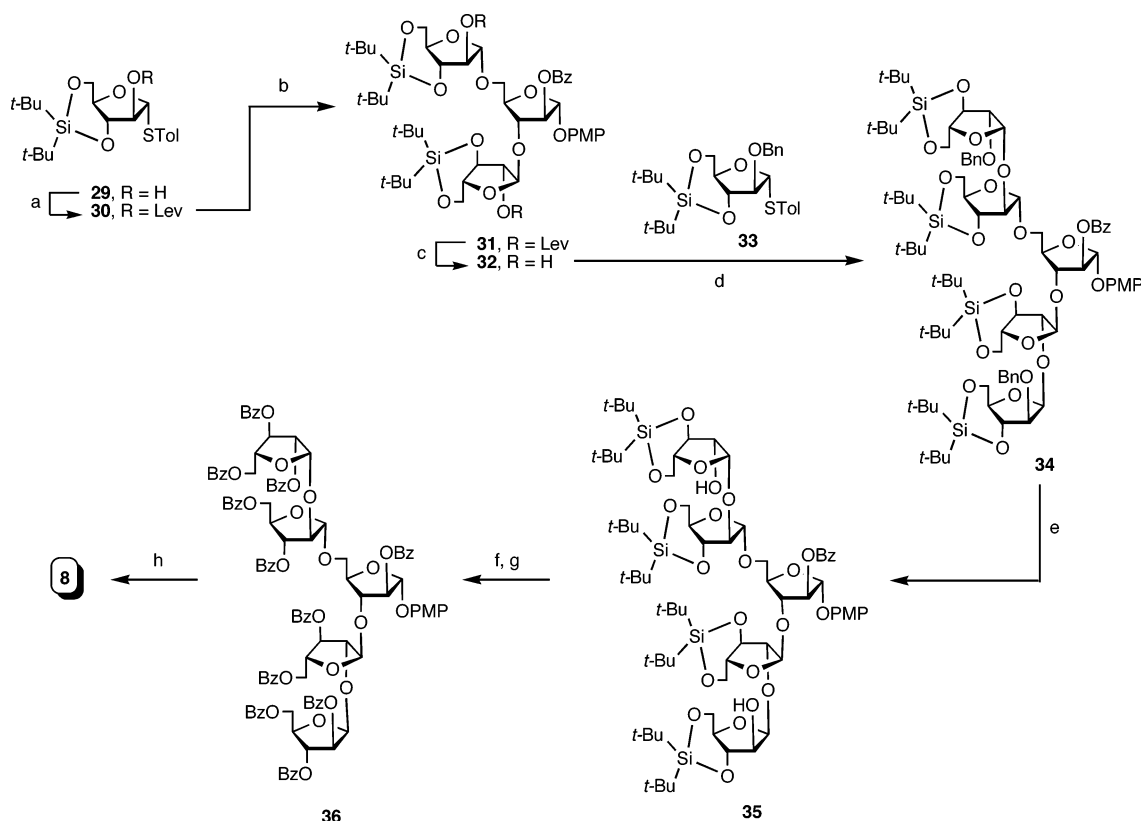
acetal were also clearly resolved in the ^1H NMR spectrum appearing at 5.54 and 4.97 ppm as small doublets ($J = 2.2$ and 2.0 Hz, respectively). These J values are slightly larger than those normally observed for α -arabinofuranosides. We ascribe this difference to the cyclic protecting group, which rigidifies the conformation of the five-membered ring, relative to those that are not fused to cyclic structures. The two levulinate esters were then cleaved with hydrazine acetate to afford, in 95% yield, diol **32**, the substrate for the key β -arabinofuranosylation reaction.

Reaction of **32** with an excess of donor thioglycoside **33**^{41,42} and NIS–AgOTf proceeded smoothly to give pentasaccharide **34**, together with a small amount (<8%) of other isomers, which could not be separated. In the ^{13}C NMR spectrum of **34**, three resonances for the anomeric carbons of the α -linked residues appeared at 107.3, 106.5, and 105.1 ppm, which could be correlated (by HMQC) with three signals in the ^1H NMR spectrum at 5.32 ppm (doublet, $J = 3.1$ Hz), 5.03 ppm (doublet, $J = 2.7$ Hz), and 5.71 ppm (singlet), respectively. In addition, the ^1H NMR spectrum of **34** indicated two β -Araf anomeric hydrogens at 5.20 ppm (doublet, $J = 4.8$ Hz) and 4.97 ppm (doublet, $J = 4.8$ Hz) that were correlated to resonances in the ^{13}C NMR spectrum at 99.5 and 99.7 ppm, respectively. As was seen with **31**, the $J_{1,2}$ values are very slightly larger for the rings with the silyl acetal protection than for rings that are not conformationally constrained. These data are all consistent^{32,39–41} with the structure of the major isomer of **34** being that shown in Scheme 4.

Having carried out the key β -arabinofuranosylation reaction, pentasaccharide **8** could be obtained by manipulation of protecting groups. Thus, removal of the benzyl groups was done by treatment with hydrogen gas and 10% palladium on carbon, which provided **35** in 84% yield. At this stage it was possible to separate the desired product **35** from the small amount of other isomers produced in the glycosylation reaction between **32** and **33**. The four silyl acetals in **35** were then cleaved by treatment with HF–pyridine to give a product that was not characterized, but instead treated with benzoyl chloride and pyridine. The result of this two-step transformation was the fully benzoylated pentasaccharide **36**, which was obtained in 70% yield. Hydrolysis of the PMP glycoside in **36** was achieved as done previously for the preparation of **6** and **7**. In the present case, **8** was obtained in 79% yield from **36** as a 1:1 mixture of isomers.

Assembly of Octadecasaccharide 3. Octadecasaccharide **3** could be produced from building blocks **5–7** without difficulty as illustrated in Scheme 5. First, heptasaccharide lactol **6** was treated with trichloroacetonitrile and DBU to provide the corresponding imidate that was quickly purified following its

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Scheme 4^a

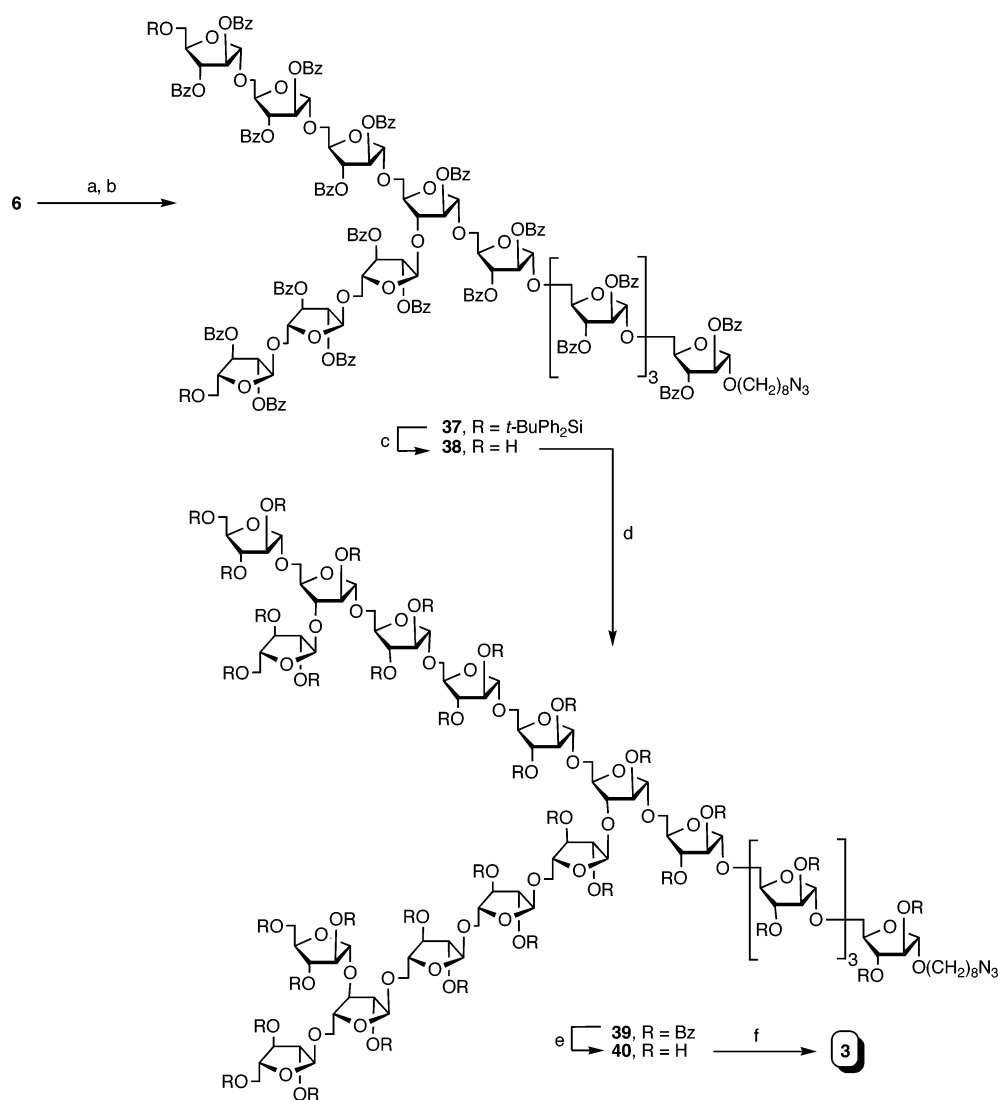
^a Reagents and conditions: (a) levulinic acid, DCC, DMAP, CH₂Cl₂, rt, 90%; (b) **24**, NIS, AgOTf, CH₂Cl₂, 0 °C, 76%; (c) H₂NNH₂-HOAc, THF, rt, 95%; (d) NIS, AgOTf, CH₂Cl₂, -30 °C, 70%; (e) 10% Pd-C, H₂, EtOAc, rt, 84%; (f) HF-pyridine, THF, 0 °C → rt; (g) BzCl, pyridine, 0 °C, 70% over two steps; (h) CAN, THF-H₂O, 0 °C, 79%.

formation, but not characterized. Instead, this donor was immediately reacted with pentasaccharide alcohol **5** and TMS-OTf, which gave a 77% yield of dodecasaccharide **37**. Given the size of this molecule and the presence of only α -Araf monosaccharide residues, there was, as expected, significant overlap in the ¹H and ¹³C spectra of **37**. That a dodecasaccharide had been produced could be ascertained from the electrospray mass spectrum, which showed a peak at $m/z = 4651$, which corresponds to the sodium adduct of **37** (C₂₆₁H₂₄₁N₃O₇₃Si₂Na). Although the donor in the glycosylation reaction leading to **37** had a participating group at C-2, which we expected would provide only the α -glycoside, it was necessary to establish this. This was possible by inspection of the ¹³C NMR spectrum, which indicated that all of the anomeric carbon resonances were between 105 and 106 ppm. Had the β -glycoside been formed, a resonance at higher field (100–105 ppm) would have been expected.³²

Conversion of the fully protected dodecasaccharide **37** into the diol required for the synthesis of the octadecasaccharide was achieved under the usual HF-pyridine conditions, which provided the product **38** in 83% yield. With **38** in hand, it was converted to the fully protected octadecasaccharide **39** in 77% yield after reaction with an excess of the trichloroacetimidate derived from trisaccharide **7**. As was done for the dodecasaccharide, a combination of mass spectrometry and ¹³C NMR spectroscopy could be used to establish the product structure ($m/z = 6425.0$ for C₃₅₇H₃₀₉N₃O₁₁₀Na⁺ and all anomeric carbon resonances between 105 and 106 ppm). Deprotection of **39** could be accomplished without incident by stirring with sodium methoxide for 36 h. Following neutralization and concentration

of the reaction mixture, purification was achieved by dissolving the residue in water and extraction with organic solvent to remove methyl benzoate; lyophilization of the aqueous layer gave a quantitative yield of **40**. Conversion of this Araf₁₈ azide species into the corresponding amine was carried out by hydrogenation (H₂, Pd/C, methanol, water), which afforded **3** in 87% yield. High-resolution electrospray mass spectrometry of **3** revealed $m/z = 2544.8972$ for the corresponding sodium adduct, as compared to the calculated $m/z = 2544.8965$. In the ¹³C NMR spectrum of **3**, all of the anomeric carbon resonances could be found clustered between 108 and 109 ppm as would be expected given the presence of only α -Araf residues (Figure 3A).

Assembly of Docosanasaccharide 4. The dodecasaccharide diol intermediate **38** employed in the synthesis of octadecasaccharide **3** was also used in the preparation of docosanasaccharide **4** (Scheme 6). Like the synthesis of **3**, this was straightforwardly done. Thus, reaction of pentasaccharide **8** with DBU and trichloroacetimidate afforded the expected trichloroacetimidate donor that was, without purification, reacted with **38** and TMS-OTf to provide the desired docosanasaccharide **41** in 77% yield. Characterization of the product by MALDI-TOF mass spectrometry showed that the compound had the expected mass ($m/z = 7785.0$ for C₄₃₃H₃₇₃N₃O₁₃₄Na). The presence of both α -Araf and β -Araf linkages was apparent from the ¹³C NMR spectrum, which showed anomeric carbon resonances not only between 105 and 107 ppm (α -Araf residues), but also around 100 ppm (β -Araf residues). Removal of the benzoyl groups as described for **39** afforded the deprotected azido docosanasaccharide **42**,

Scheme 5^a

^a Reagents and conditions: (a) CCl₃CN, DBU, CH₂Cl₂, 0 °C → rt; (b) **5**, TMSOTf, CH₂Cl₂, -30 → -5 °C, 77% over two steps; (c) HF–pyridine, THF, 0 °C → rt, 83%; (d) **7**, CCl₃CN, DBU, CH₂Cl₂, 0 °C → rt, then TMSOTf, CH₂Cl₂, -30 → -5 °C, 77% over two steps; (e) NaOCH₃, CH₃OH–CH₂Cl₂, rt, quantitative; (f) 10% Pd–C, H₂, CH₃OH–H₂O, rt, 87%.

which was subsequently reduced by hydrogenation. The target **4** was obtained in 89% yield in two steps from **41**.

The high-resolution electrospray mass spectrum of docosanasaccharide **4** showed a molecular ion of the sodium adduct at $m/z = 3073.0655$, which was identical to the calculated exact mass for this molecule. In the 125 MHz ¹³C NMR spectrum (Figure 3B), the majority of the anomeric carbon signals were, as in the case of **3**, located between 108 and 109 ppm. These resonances arise from 14 of the α-Araf residues and were not sufficiently resolved for assignment. Although at this field strength the signals arising from the remaining eight anomeric carbons did not fully resolve, appearing instead as four resonances, they provide further important support for the structure of **4**. Two of these signals, corresponding to the four β-Araf moieties in **4**, were present at 101.5 and 101.6 ppm. Two peaks were also present at 106.3 and 106.5 ppm, which, on the basis of previous assignments,⁴³ we assign as the four

α-Araf residues that bear the β-Araf moieties on O-2. We note that these chemical shifts and doubling of resonances are consistent with those reported for the arabinan domain of AG in a recent solid-state NMR study of live mycobacteria.⁴⁴

Conclusions

In summary, we report here the first total synthesis of the 22-residue arabinan domain of mycobacterial arabinogalactan and a proposed octadecasaccharide biosynthetic precursor, both functionalized with an aminooctyl linker arm. Key features of the route to the targets **3** and **4** were the preparation of tri- to heptasaccharide building blocks possessing only benzoyl protection that were coupled in a highly convergent manner via glycosyl trichloroacetimidate donors. Once the four building blocks were in hand, each of the targets could be obtained in only six steps. This route produced 11 and 17 mg quantities of **3** and **4**, respectively, which will be useful tools in probing mycobacterial arabinan biosynthesis, a critical prerequisite for

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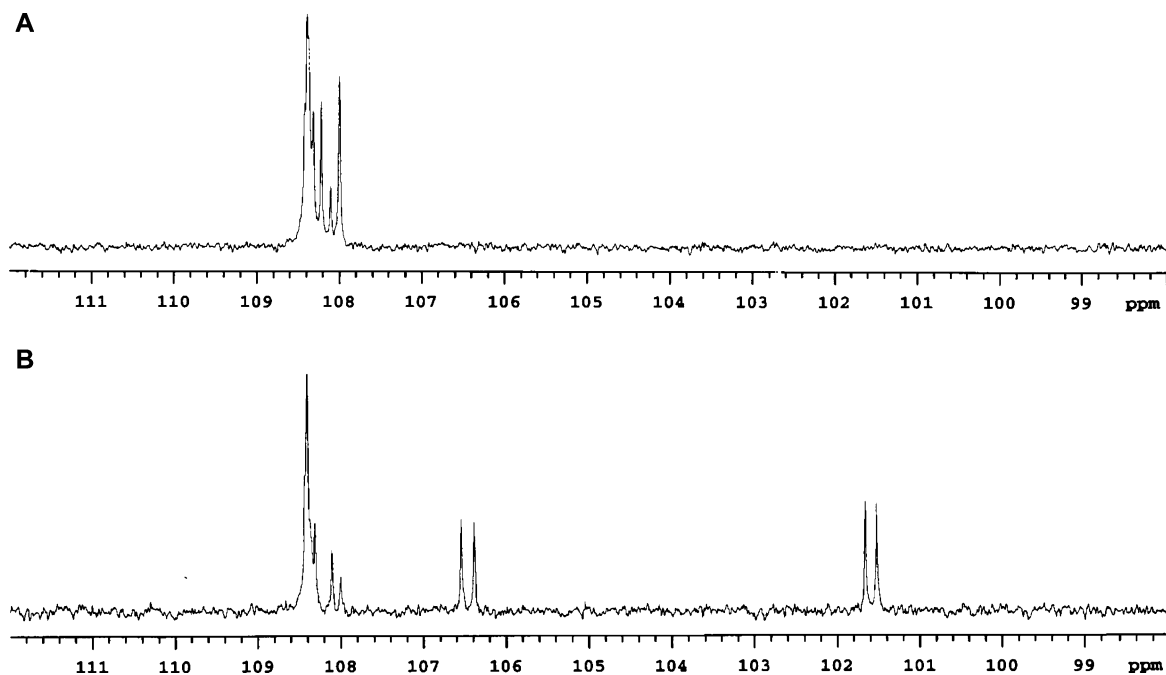
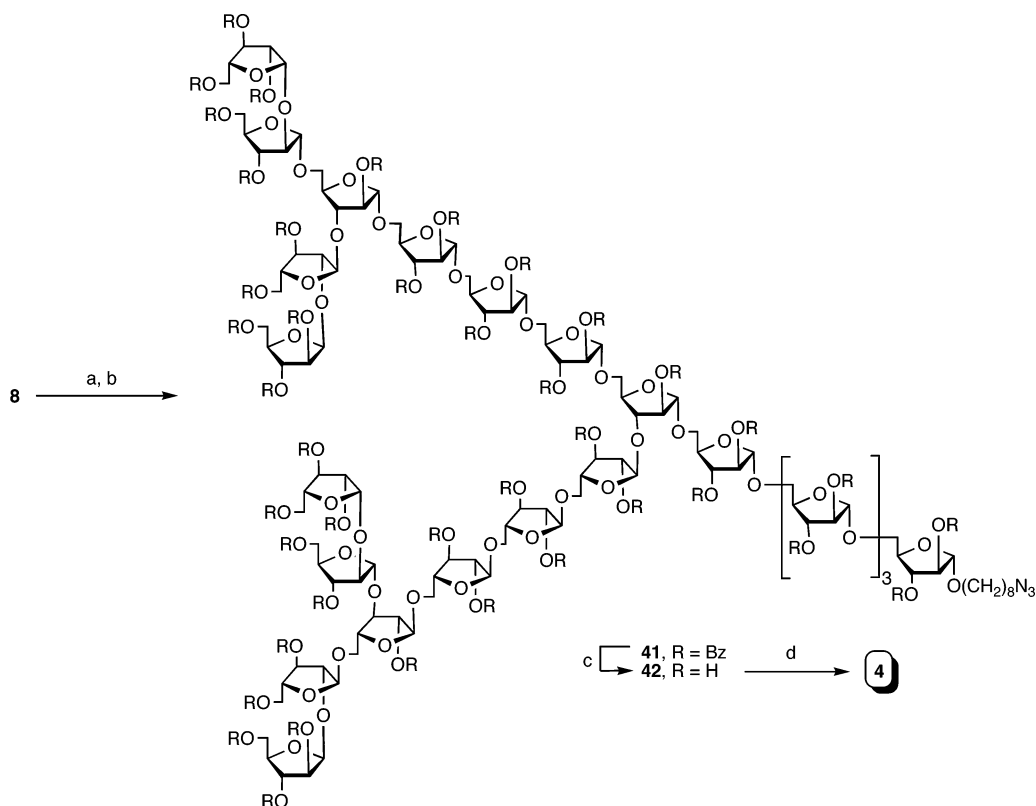


Figure 3. Partial ^{13}C NMR spectra of **3** and **4**. (A) Anomeric carbon region for **3**. As expected for the molecule, which contains only α -Araf residues, all signals are clustered between 108 and 109 ppm. (B) Anomeric carbon region for **4**. The presence of the β -Araf residues is clearly apparent from the resonances around 101.5 ppm; the resonances around 106.5 ppm are proposed to arise from the anomeric carbons of the four α -Araf moieties that are glycosylated by β -Araf on O-2.

Scheme 6^a



^a Reagents and conditions: (a) CCl_3CN , DBU, CH_2Cl_2 , $0\text{ }^\circ\text{C} \rightarrow \text{rt}$; (b) **38**, TMSOTf, CH_2Cl_2 , $-30 \rightarrow -5\text{ }^\circ\text{C}$, 77% over two steps; (c) NaOCH_3 , $\text{CH}_3\text{OH}-\text{CH}_2\text{Cl}_2$, rt, quantitative; (d) 10% Pd-C, H_2 , $\text{CH}_3\text{OH}-\text{H}_2\text{O}$, rt, 89%.

the identification of novel anti-TB agents targeting these biochemical pathways. In addition, we envisage that **4** will be a valuable substrate for the mycolyltransferases that add mycolic acids to the AG.⁴⁵

Experimental Section

General Methods. Reactions were carried out in oven-dried glassware. All reagents used were purchased from commercial sources and were used without further purification unless noted. Reaction

81.5(9), 81.5(6), 77.6, 77.3, 77.2, 67.3 (octyl OCH₂), 66.1 (C-5), 66.0 (C-5), 65.8(6) (C-5), 65.8(3) (C-5), 62.3 (C-5), 51.4 (octyl CH₂N₃), 29.4 (octyl CH₂), 29.2 (octyl CH₂), 29.0 (octyl CH₂), 28.8 (octyl CH₂), 26.6 (octyl CH₂), 26.0 (octyl CH₂); ESIMS *m/z* calcd for (M + Na) C₁₀₃H₇₇N₃O₃₁ 1894.5999, found 1894.5998.

2,3-Di-*O*-benzoyl-5-*O*-(*tert*-butyldiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3-di-*O*-benzoyl-5-*O*-(*tert*-butyldiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-*O*-benzoyl-D-arabinofuranose (6). To a solution of compound **27** (1.85 g, 0.64 mmol) in THF–H₂O (4:1, 90 mL) at 0 °C was added CAN (8.8 g, 16.05 mmol), and the reaction mixture was stirred for 20 min. The reaction mixture was diluted with EtOAc (200 mL) and brine (250 mL) and stirred well. The EtOAc layer was separated, and the aqueous phase was extracted with EtOAc (75 mL \times 2). The combined organic layer was washed with water (75 mL), a saturated aqueous NaHCO₃ solution (150 mL), and water (75 mL), dried (Na₂SO₄), and concentrated to give a crude that was purified by column chromatography (7:3 hexanes–EtOAc) to afford **6** (1.43 g, 1:1 α/β mixture, 80%) as a syrup. About 5% of the starting material was also recovered. Data for **6**: *R*_f 0.20 (65:35 hexanes–EtOAc); ¹H NMR (600 MHz, CDCl₃, δ _H) 8.15–7.85 (m, 26 H, Ar), 7.73–7.65 (m, 8 H, Ar), 7.64–7.20 (m, 51 H, Ar), 5.98–5.90 (m, 0.5 H), 5.83–5.70 (m, 1.5 H), 5.68–5.60 (m, 7 H), 5.58–5.55 (m, 5 H), 5.42–5.27 (m, 6 H), 4.66–4.56 (m, 3 H), 4.52–4.46 (m, 4 H), 4.24–4.14 (m, 5 H), 4.08–3.80 (m, 10 H), 3.67–3.63 (m, 1H), 1.01 (s, 18 H, C(CH₃)₃ \times 2); ¹³C NMR (125 MHz, CDCl₃, δ _C) 166.1 (C=O), 165.9(1) (C=O), 165.9 (C=O), 165.6(7) (C=O), 165.6 (C=O), 165.5(7) (C=O), 165.5(3) (C=O), 165.5(2) (C=O), 165.4(9) (C=O), 165.4(5) (C=O), 165.3(6) (C=O), 165.3(4) (C=O), 165.2(C=O), 165.1(7) (C=O), 165.1 (C=O), 165.0(9) (C=O), 165.0(7) (C=O), 165.0(4) (C=O), 135.6(6) (Ar), 135.6(4) (Ar), 133.6 (Ar), 133.5 (Ar), 133.4 (Ar), 133.3(9) (Ar), 133.3(7) (Ar), 133.3(4) (Ar), 133.2(9) (Ar), 133.2(4) (Ar), 133.2(1) (Ar), 133.1 (Ar), 132.9 (Ar), 132.3 (Ar), 129.9 (Ar), 129.8(8) (Ar), 129.8(1) (Ar), 129.7(6) (Ar), 129.7(2) (Ar), 129.6 (Ar), 129.3 (Ar), 129.2 (Ar), 129.1(5) (Ar), 129.1(3) (Ar), 129.0 (Ar), 128.9(9) (Ar), 128.9(6) (Ar), 128.8(8) (Ar), 128.8 (Ar), 128.5 (Ar), 128.4(7) (Ar), 128.4(1) (Ar), 128.3 (Ar), 128.2- (5) (Ar), 128.2(3) (Ar), 128.1 (Ar), 127.6 (Ar), 106.4 (C-1), 106.1 (C-1), 105.9(9) (C-1), 105.9(7) (C-1), 105.9(1) (C-1), 105.8(6) (C-1), 105.8 (C-1), 105.1 (C-1), 103.4 (C-1), 101.1 (C-1), 94.9 (C-1), 84.2, 83.1, 82.8, 82.6(6), 82.6(1), 82.4, 82.3, 82.2, 82.1, 81.9(8), 81.9(4), 81.8, 81.6, 81.5(7), 81.5(2), 81.5, 81.4, 80.1, 79.5, 79.3, 79.1, 78.2, 77.3, 77.2, 76.6, 66.6 (C-5), 65.8 (C-5), 65.7(8) (C-5), 65.7(2) (C-5), 65.6 (C-5), 65.5 (C-5), 63.4 (C-5), 60.3 (C-5), 26.7 (C(CH₃)₃), 26.6 (C(CH₃)₃) 21.0 (C(CH₃)₃), 19.2 (C(CH₃)₃); MALDI-TOFMS *m/z* calcd for (M + Na) C₁₅₈H₁₄₆O₄₂Si₂ 2794.7, found: 2794.1

2,3,5-Tri-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3,5-tri-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-*O*-benzoyl-D-arabinofuranose (7). Compound **7** was prepared as a syrup (0.47 g, 1:1 α/β ratio, 85%) from compound **28** (0.60 g, 0.48 mmol) in THF–H₂O (4:1, 30 mL) and CAN (6.58 g, 12.0 mmol) as described for **6**: *R*_f 0.27 (65:35 hexanes–EtOAc); ¹H NMR (400 MHz, CDCl₃, δ _H) 8.10–7.90 (m, 14 H, Ar), 7.62–7.20 (m, 21 H, Ar), 5.72 (d, 0.5 H, *J* = 4.4 Hz), 5.65–5.52 (m, 6 H), 5.45–5.42 (m, 1 H), 5.36 (s, 0.5 H), 5.32 (dd, 0.5 H, *J* = 6.6, 4.5 Hz), 4.91 (dd, 0.5 H, *J* = 6.5, 6.5 Hz), 4.83–4.56 (m, 7.5 H), 4.20 (ddd, 0.5 H, *J* = 9.2, 6.2, 2.9 Hz), 4.14–4.06 (m, 1 H), 3.95 (dd, 0.5 H, *J* = 11.8, 2.7 Hz), 3.88 (dd, 0.5 H, *J* = 11.2, 3.4 Hz); ¹³C NMR (125 MHz, CDCl₃, δ _C) 166.2 (C=O), 166.1 (C=O), 166.0(4) (C=O), 166.0(2) (C=O), 165.9(4) (C=O), 165.9(1) (C=O), 165.7 (C=O), 165.6(5) (C=O), 165.6(3) (C=O), 165.3 (C=O), 165.2(4) (C=O), 165.2(3) (C=O), 133.6(9) (Ar), 133.6(5) (Ar), 133.5(9) (Ar), 133.5(6) (Ar), 133.5(1) (Ar), 133.4(9) (Ar), 133.4(4) (Ar), 133.4(2) (Ar), 133.2(Ar), 133.0(8) (Ar), 133.0(2) (Ar), 129.9(8) (Ar), 129.9(4) (Ar), 129.9(2) (Ar), 129.9(0) (Ar), 129.8(7) (Ar), 129.8(4) (Ar), 129.7- (6) (Ar), 129.7(3) (Ar), 129.7(0) (Ar), 129.6(Ar), 129.5(Ar), 129.2-

(Ar), 129.0(3) (Ar), 129.0(1) (Ar), 128.9(8) (Ar), 128.9(5) (Ar), 128.8(9) (Ar), 128.8(6) (Ar), 128.7 (Ar), 128.5(6) (Ar), 128.5(3) (Ar), 128.5(0) (Ar), 128.4(7) (Ar), 128.4(3) (Ar), 128.3(9) (Ar), 128.3(5) (Ar), 128.3- (3) (Ar), 128.3(2) (Ar), 128.2(Ar), 106.5 (C-1), 106.1 (C-1), 106.0 (C-1), 105.4 (C-1), 101.0 (C-1), 94.9 (C-1), 82.3, 82.2, 82.1(6), 82.1(1), 82.0, 81.7, 81.6, 81.5, 81.4, 81.1, 80.4, 79.1, 78.9, 78.3, 77.7, 77.6, 77.5, 66.6 (C-5), 66.3 (C-5), 63.7 (C-5 \times 3), 63.5 (C-5); ESIMS *m/z* calcd for (M + Na) C₆₄H₅₄O₂₀ 1165.3104, found 1165.3100.

2,3,5-Tri-*O*-benzoyl- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3,5-tri-*O*-benzoyl- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-di-*O*-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-*O*-benzoyl-D-arabinofuranose (8). Compound **8** was prepared as a syrup (0.486 g, 1:1 α/β mixture, 79%) from compound **36** (0.65 g, 0.336 mmol) in THF–H₂O (4:1, 35 mL) and CAN (4.61 g, 8.4 mmol) as described for **6**. About 6% of the starting material was also recovered. Data for **8**: *R*_f 0.22 (65:35 hexanes–EtOAc); ¹H NMR (500 MHz, CDCl₃, δ _H) 8.10–7.80 (m, 23 H, Ar), 7.62–7.18 (m, 36 H, Ar), 6.02–5.88 (m, 2 H), 5.86–5.75 (m, 1 H), 5.74–5.62 (m, 1 H), 5.60–5.49 (m, 1.5 H), 5.48–5.32 (m, 4 H), 5.20 (d, 0.5 H, *J* = 1.7 Hz), 5.13 (s, 0.5 H), 5.10–5.05 (m, 1 H), 4.82–4.30 (m, 11 H), 4.25–4.10 (m, 1.5 H), 4.01–3.88 (m, 1.5 H), 3.78–3.70 (m, 1.5 H); ¹³C NMR (125 MHz, CDCl₃, δ _C) 166.4 (C=O), 166.1 (C=O), 165.9 (C=O), 165.8 (C=O), 165.7 (C=O), 165.6 (C=O), 165.5 (C=O), 165.4 (C=O), 165.3 (C=O), 133.6 (Ar), 133.5 (Ar), 133.4(8) (Ar), 133.4(3) (Ar), 133.3 (Ar), 133.2(7) (Ar), 133.2(3) (Ar), 133.1 (Ar), 132.8(8) (Ar), 132.8(2) (Ar), 129.9 (Ar), 129.8 (Ar), 129.7(9) (Ar), 129.7(3) (Ar), 129.6(9) (Ar), 129.6(3) (Ar), 129.5 (Ar), 129.2 (Ar), 129.1(8) (Ar), 129.1(2) (Ar), 129.0 (Ar), 128.9(9) (Ar), 128.9(5) (Ar), 128.7 (Ar), 128.6 (Ar), 128.5 (Ar), 128.4(8) (Ar), 128.4(4) (Ar), 128.3(7) (Ar), 128.3(1) (Ar), 128.2 (Ar), 128.1 (Ar), 106.5 (C-1), 105.9 (C-1), 104.9 (C-1), 104.8 (C-1), 100.9 (C-1), 100.5 (C-1), 100.3 (C-1), 100.2 (C-1), 100.1 (C-1), 94.7 (C-1), 85.5, 85.2, 84.9, 84.7, 82.9, 81.7, 80.9, 80.8, 80.5, 80.3, 79.4- (5), 79.4(0), 79.2, 79.0, 78.2, 78.0, 77.9, 77.8, 77.7, 77.6, 77.5, 77.3, 76.5, 76.4, 76.3, 66.5 (C-5), 65.7 (C-5), 65.6 (C-5), 64.3 (C-5), 64.2 (C-5), 64.1 (C-5); ESIMS *m/z* calcd for (M + Na) C₁₀₂H₈₆O₃₂ 1845.4996, found 1845.4999.

***p*-Tolyl 2,3-Di-*O*-benzoyl-5-*O*-(*tert*-butyldiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-*O*-benzoyl-1-thio- α -D-arabinofuranoside (15).** To a solution of alcohol **12** (0.6 g, 1.0 mmol) and trichloroacetimidate (0.5 mL, 5 mmol) in CH₂Cl₂ (9 mL) at 0 °C was added DBU (15 μ L, 0.1 mmol). The reaction mixture was stirred at 0 °C for 30 min and then warmed to room temperature over 30 min. The solvent was then removed under vacuum, and a solution of dry hexane–toluene (2:3, 10 mL) was added. After being stirred for 5 min, this solution was quickly filtered through a short column of silica gel and Na₂SO₄ (~1:1). The resulting solution was then concentrated to yield the trichloroacetimidate derivative **13**, which was used without any further purification. Alternatively, the syrupy residue obtained after the initial solvent evaporation following the reaction could be quickly filtered through silica gel (4:1 hexanes–EtOAc). The fractions containing the trichloroacetimidate derivative were concentrated and used immediately without any further purification. The trichloroacetimidate derivative **13** in CH₂Cl₂ (6 mL) was added to a solution of alcohol **14**³³ (0.39 g, 0.84 mmol) in CH₂Cl₂ (26 mL) containing 4 Å molecular sieves (0.4 g; stirred already for about 30 min) at –30 °C. A solution of TMSOTf (72 μ L, 0.4 mmol) in CH₂Cl₂ (0.72 mL) was added dropwise over a period of 5 min. The reaction mixture was then warmed to –5 °C over a period of ~20 min and quenched by the addition of Et₃N (0.1 mL). The solution was diluted with CH₂Cl₂ (15 mL) and filtered. The filtrate was concentrated to a syrup that was purified by column chromatography (9:1 hexanes–EtOAc) to afford **15** (0.67 g, 76% over two steps) as a syrup: *R*_f 0.42 (4:1 hexanes–EtOAc); [α]_D +40.5 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ _H) 8.14–7.92 (m, 8 H, Ar), 7.76–7.70 (m, 4 H, Ar), 7.63–7.53 (m, 2 H, Ar), 7.50–7.22 (m, 18 H, Ar), 7.15–7.05 (m, 2 H, Ar), 5.76–5.71 (m, 3 H, H-1, H-2, H-3), 5.65 (d, 1 H, *J* = 5.0 Hz, H-3'), 5.57 (d, 1 H, *J* = 1.3 Hz, H-2'),

5.38 (s, 1 H, H-1'), 4.74 (dd, 1 H, $J = 7.5, 4.1$ Hz, H-4), 4.52 (dd, 1 H, $J = 9.3, 4.6$ Hz, H-4'), 4.24 (dd, 1 H, $J = 11.3, 4.4$ Hz, H-5), 4.22–3.94 (m, 3 H, H-5, H-5' $\times 2$), 2.30 (s, 3 H, Ar-CH₃), 1.04 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.5 (C=O $\times 2$), 165.2–(7) (C=O), 165.2(2) (C=O), 137.8 (Ar), 135.6(7) (Ar), 135.6(5) (Ar), 133.4(8) (Ar), 133.4(2) (Ar), 133.3(Ar), 133.2(7) (Ar), 133.2(2) (Ar), 133.0(Ar), 132.6 (Ar), 130.0 (Ar), 129.9 (Ar), 129.8 (Ar), 129.6 (Ar), 129.2(8) (Ar), 129.2(2) (Ar), 128.9(Ar), 128.5 (Ar), 128.4 (Ar), 128.3 (Ar), 128.2 (Ar), 127.6 (Ar), 106.0 (C-1'), 91.5 (C-1), 83.2 (C-4'), 82.1, 82.0 ($\times 2$), 77.5 (C-3), 77.3 (C-3'), 65.8 (C-5), 63.4 (C-5'), 26.7 (C(CH₃)₃), 21.1 (Ar-CH₃), 19.3 (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₆₁H₅₈O₁₂Si 1065.3308, found 1065.3310.

8-Azido-octyl 2,3-Di-O-benzoyl-5-O-(tert-butyl)diphenylsilyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranoside (16). Thioglycoside **15** (2.0 g, 1.92 mmol) and alcohol **11** (0.85 g, 1.67 mmol) were dried over P₂O₅ under vacuum for 6 h and then dissolved in CH₂Cl₂ (60 mL), and the resulting solution was cooled to 0 °C. Powdered 4 Å molecular sieves (0.5 g) were added, and the suspension was stirred for 30 min at 0 °C before *N*-iodosuccinimide (0.52 g, 2.3 mmol) and silver triflate (0.1 g, 0.38 mmol) were added. The reaction mixture was stirred for 20 min, neutralized with Et₃N, diluted with CH₂Cl₂ (30 mL), and filtered through Celite. The filtrate was washed successively with a saturated aqueous Na₂S₂O₃ solution (75 mL $\times 2$) and water (75 mL) before being dried (Na₂SO₄) and concentrated. The crude residue was purified by column chromatography (6:1 hexanes–EtOAc) to afford **16** (2.0 g, 84%) as a syrup: R_f 0.34 (4:1 hexanes–EtOAc); $[\alpha]_D +5.4$ (c 1.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.10–7.85 (m, 13 H, Ar), 7.75–7.65 (m, 4 H, Ar), 7.60–7.20 (m, 23 H, Ar), 5.66–5.62 (m, 4 H), 5.57 (s, 1 H), 5.51 (s, 1 H), 5.40 (s, 1 H), 5.39 (s, 1 H), 5.21 (s, 1 H), 4.63 (dd, 1 H, $J = 7.7, 4.5$ Hz), 4.50 (dd, 1 H, $J = 8.4, 4.1$ Hz), 4.44 (dd, 1 H, $J = 7.6, 4.6$ Hz), 4.24–4.17 (m, 2 H), 4.0–3.91 (m, 4 H), 3.75 (ddd, 1 H, $J = 13.3, 9.4, 6.7$ Hz, octyl OCH₂), 3.50 (ddd, 1 H, $J = 10.9, 9.4, 6.2$ Hz, octyl OCH₂), 3.22 (dd, 2 H, $J = 6.9, 6.9$ Hz, octyl CH₂N₃), 1.70–1.50 (m, 4 H, octyl CH₂), 1.42–1.22 (m, 8 H, octyl CH₂), 1.02 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.6(3) (C=O), 165.6 (C=O), 165.4(5) (C=O), 165.4 (C=O), 165.1(8) (C=O), 165.1 (C=O), 135.6(6) (Ar), 135.6 (Ar), 133.3–(6) (Ar), 133.3(4) (Ar), 133.3(1) (Ar), 133.2(3) (Ar), 133.2(1) (Ar), 133.1 (Ar), 133.0 (Ar), 129.9 (Ar), 129.8(7) (Ar), 129.8(3) (Ar), 129.8 (Ar), 129.7 (Ar), 129.6 (Ar), 129.3(8) (Ar), 129.3(1) (Ar), 129.2(8) (Ar), 129.2 (Ar), 129.1 (Ar), 129.0 (Ar), 128.4(9) (Ar), 128.4(6) (Ar), 128.4(2) (Ar), 128.3(9) (Ar), 128.3(3) (Ar), 128.2(9) (Ar), 128.2(4) (Ar), 128.1 (Ar), 127.6 (Ar), 105.9 (C-1), 105.8 (C-1), 105.5 (C-1), 83.1, 82.1(4), 82.1(2), 81.8, 81.7, 81.5, 77.3(6), 77.3(3), 67.3, 66.0 (C-5), 65.8 (C-5), 63.4 (C-5), 51.4 (octyl CH₂N₃), 29.4 (octyl CH₂), 29.2 (octyl CH₂), 29.1 (octyl CH₂), 28.8 (octyl CH₂), 26.7 (C(CH₃)₃), 26.6 (octyl CH₂), 26.0 (octyl CH₂), 19.2 (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₈₁H₈₃N₃O₁₉Si 1452.5280, found 1452.5282.

8-Azido-octyl 2,3-Di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranoside (17). Compound **17** was prepared as a syrup (1.55 g, 95%) from compound **16** (1.96 g, 1.37 mmol) and HF–pyridine (1.0 mL) in pyridine–THF (1:5, 30 mL) as described for compound **5**: R_f 0.13 (3:1 hexanes–EtOAc); $[\alpha]_D +1.4$ (c 1.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.10–7.98 (m, 8 H), 7.98–7.90 (m, 4 H), 7.61–7.32 (m, 14 H), 7.20–7.30 (m, 4 H), 5.67–5.61 (m, 4 H), 5.51 (s, 1 H), 5.44–5.39 (m, 3 H), 5.22 (s, 1 H), 4.63 (dd, 1 H, $J = 7.7, 4.5$ Hz), 4.47 (dd, 1 H, $J = 4.1, 8.4$ Hz), 4.43 (dd, 1 H, $J = 7.6, 4.6$ Hz), 4.23–4.16 (m, 2 H), 4.04–3.90 (m, 4 H), 3.75 (ddd, 1 H, $J = 13.3, 9.4, 6.7$ Hz, octyl OCH₂), 3.50 (ddd, 1 H, $J = 10.9, 9.4, 6.2$ Hz, octyl OCH₂), 3.21 (dd, 2 H, $J = 6.9, 6.9$ Hz, octyl CH₂N₃), 1.70–1.50 (m, 4 H, octyl CH₂), 1.50–1.20 (m, 8 H, octyl CH₂); ¹³C NMR (125 MHz, CDCl₃, δ_c) 166.0 (C=O), 165.6(7) (C=O), 165.6(6) (C=O), 165.4 (C=O), 165.1 (C=O), 165.0 (C=O), 133.4(9) (Ar), 133.4(1) (Ar), 133.3 (Ar), 133.2 (Ar), 133.1 (Ar), 129.8(7) (Ar), 129.8(5) (Ar), 129.8 (Ar),

129.7 (Ar), 129.3 (Ar), 129.1(8) (Ar), 129.1 (Ar), 129.0 (Ar), 128.9 (Ar), 128.4(9) (Ar), 128.4(1) (Ar), 128.2(9) (Ar), 128.2(5) (Ar), 105.8–(4) (C-1), 105.8(0) (C-1), 105.5 (C-1), 83.6, 82.0, 81.8, 81.7, 81.6(9), 81.6, 77.7, 77.3(7), 77.3(3), 67.3 (octyl OCH₂), 66.1 (C-5), 66.0 (C-5), 62.3 (C-5), 51.4 (octyl CH₂N₃), 29.4 (octyl CH₂), 29.2 (octyl CH₂), 29.0 (octyl CH₂), 28.8 (octyl CH₂), 26.6 (octyl CH₂), 26.0 (octyl CH₂); ESIMS m/z calcd for (M + Na) C₆₅H₆₅N₃O₁₉ 1214.4103, found 1214.4104.

8-Azido-octyl 2,3-Di-O-benzoyl-5-O-(tert-butyl)diphenylsilyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranoside (18). Compound **18** was prepared as a syrup (2.32 g, 86%) from thioglycoside **15** (1.53 g, 1.47 mmol), alcohol **17** (1.52 g, 1.28 mmol), 4 Å molecular sieves (0.5 g), *N*-iodosuccinimide (0.4 g, 1.76 mmol), and silver triflate (0.075 g, 0.29 mmol) in CH₂Cl₂ (60 mL) as described for **16**: R_f 0.22 (3:1 hexanes–EtOAc); $[\alpha]_D +11.6$ (c 1.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.10–7.85 (m, 19 H, Ar), 7.75–7.63 (m, 4 H, Ar), 7.60–7.20 (m, 37 H, Ar), 5.68–5.60 (m, 9 H), 5.56 (s, 1 H), 5.51(s, 1 H), 5.41–5.39 (m, 3 H), 5.37 (s, 1 H), 5.21(s, 1 H), 4.64–4.57 (m, 3 H), 4.49 (dd, 1 H, $J = 9.2, 4.5$ Hz), 4.43 (dd, 1 H, $J = 7.1, 4.2$ Hz), 4.23–4.14 (m, 4 H), 4.00–3.88 (m, 5 H), 3.75 (ddd, 1 H, $J = 13.5, 9.4, 6.7$ Hz, octyl OCH₂), 3.50 (ddd, 1 H, $J = 13.1, 9.4, 6.2$ Hz, octyl OCH₂), 3.22 (dd, 2 H, $J = 6.9, 6.9$ Hz, octyl CH₂N₃), 1.70–1.50 (m, 4 H, octyl CH₂), 1.50–1.23 (m, 8 H, octyl CH₂), 1.01(s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.6 (C=O), 165.5(9) (C=O), 165.5(7) (C=O), 165.5(5) (C=O), 165.4(5) (C=O), 165.4(2) (C=O), 165.1(5) (C=O), 165.1(3) (C=O), 165.1 (C=O), 165.0 (C=O), 135.6(5) (Ar), 135.6(3) (Ar), 133.3(6) (Ar), 133.3(2) (Ar), 133.2(9) (Ar), 133.2(7) (Ar), 133.2(2) (Ar), 133.2–(1) (Ar), 133.1 (Ar), 133.0(6) (Ar), 133.0(0) (Ar), 129.9 (Ar), 129.8–(8) (Ar), 129.8(2) (Ar), 129.7(Ar), 129.6 (Ar), 129.3(6) (Ar), 129.3(1) (Ar), 129.2 (Ar), 129.1 (Ar), 129.0 (Ar), 128.4(7) (Ar), 128.4 (Ar), 128.3(8) (Ar), 128.3(3) (Ar), 128.2 (Ar), 128.1 (Ar), 127.6 (Ar), 105.9 (C-1 $\times 2$), 105.8(9) (C-1), 105.8(3) (C-1), 105.5 (C-1), 83.1, 82.1(1), 82.1(0), 81.8, 81.7, 81.5(6), 81.5(1), 77.3, 77.2, 67.3 (octyl OCH₂), 66.0 (C-5), 65.8(6) (C-5), 65.8(3) (C-5 $\times 2$), 63.4 (C-5), 51.4 (octyl CH₂N₃), 29.4 (octyl CH₂), 29.2 (octyl CH₂), 29.1 (octyl CH₂), 28.8 (octyl CH₂), 26.7 (C(CH₃)₃), 26.6 (octyl CH₂), 26.0 (octyl CH₂), 19.2 (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₁₁₉H₁₁₅N₃O₃₁Si 2133.7, found 2133.7.

***p*-Methoxyphenyl 2,3-Di-O-benzoyl-5-O-(tert-butyl)diphenylsilyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3-di-O-benzoyl-5-O-(tert-butyl)diphenylsilyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (25).** Compound **25** was prepared as a syrup (0.7 g, 74%) from thioglycoside **9** (1.34 g, 1.99 mmol), acceptor alcohol **24** (0.23 g, 0.62 mmol), 4 Å molecular sieves (0.5 g), *N*-iodosuccinimide (0.45 g, 1.99 mmol), and silver triflate (0.025 g, 0.09 mmol) in CH₂Cl₂ (30 mL) as described for **16**. In addition about 8% of a monoglycosylated product was also isolated. Data for **25**: R_f 0.24 (4:1 hexanes–EtOAc); $[\alpha]_D +26.5$ (c 0.9, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.12–7.90 (m, 10 H, Ar), 7.72–7.61 (m, 8 H, Ar), 7.60–7.20 (m, 27 H, Ar), 7.05 (d, 2 H, $J = 8.9$ Hz, Ar), 6.78 (d, 2 H, $J = 8.9$ Hz, Ar), 5.77 (s, 1 H, H-1), 5.66 (s, 1 H), 5.65–5.60 (m, 3 H), 5.60 (s, 1 H, H-1), 5.50 (s, 1 H), 5.33 (s, 1 H, H-1), 4.61–4.53 (m, 2 H), 4.39 (dd, 2 H, $J = 9.2, 4.6$ Hz), 4.10 (dd, 1 H, $J = 11.4, 4.3$ Hz), 3.98–3.87 (m, 5 H), 3.73 (s, 3 H, OCH₃), 1.01 (s, 9 H, C(CH₃)₃), 0.98 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.4(7) (C=O $\times 2$), 165.4(1) (C=O), 165.1(7) (C=O), 165.1(5) (C=O), 155.0 (Ar), 150.3 (Ar), 135.6–(6) (Ar), 135.6(2) (Ar), 135.5 (Ar), 133.3 (Ar), 133.2(9) (Ar), 133.2(7) (Ar), 133.2(4) (Ar), 133.1(8) (Ar), 133.1(4) (Ar), 133.1(1) (Ar), 133.0–(5) (Ar), 130.0 (Ar), 129.9 (Ar), 129.8 (Ar), 129.7(8) (Ar), 129.7(6) (Ar), 129.6 (Ar), 129.4 (Ar), 129.3 (Ar), 129.2(7) (Ar), 129.2 (Ar), 129.1 (Ar), 128.4 (Ar), 128.3(4) (Ar), 128.3 (Ar), 127.6 (Ar), 118.5 (Ar), 114.5 (Ar), 106.2 (C-1), 105.6 (C-1), 105.3 (C-1), 84.0, 83.4, 82.8, 82.7, 82.0, 81.9, 80.7, 77.2, 77.1, 65.9 (C-5), 63.5 (C-5), 63.3

(C-5), 55.6 (OCH₃), 26.7(6) (C(CH₃)₃), 26.7(4) (C(CH₃)₃), 19.2(8) (C(CH₃)₃), 19.2(1) (C(CH₃)₃); ESIMS *m/z* calcd for (M + Na) C₈₉H₈₈O₁₉Si₂ 1539.5349, found 1539.5356.

p-Methoxyphenyl 2,3-Di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (26). Compound **26** was prepared as a syrup (1.43 g, 89%) from compound **25** (2.35 g, 1.55 mmol) and HF-pyridine (1.0 mL) in THF-pyridine (5:1, 30 mL) as described for **5**: *R_f* 0.11 (3:2 hexanes-EtOAc); [α]_D +15.6 (c 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ _H) 8.10–7.90 (m, 10 H, Ar), 7.65–7.33 (m, 13 H, Ar), 7.18–7.24 (m, 2 H, Ar), 7.05 (d, 2 H, *J* = 8.9 Hz, Ar), 6.81 (d, 2 H, *J* = 8.9 Hz, Ar), 5.77 (s, 1 H, H-1), 5.65 (s, 1 H), 5.62 (d, 2 H, *J* = 4.3 Hz), 5.35–5.30 (m, 3 H), 5.28 (d, 1 H, *J* = 4.6 Hz), 4.65 (d, 1 H, *J* = 5.7 Hz), 4.52–4.48 (m, 1 H), 4.40 (dd, 1 H, *J* = 8.3, 4.0 Hz), 4.34 (dd, 1 H, *J* = 8.4, 4.0 Hz), 4.07–4.0 (m, 2 H), 3.98–3.84 (m, 4 H), 3.76 (s, 3 H, OCH₃); ¹³C NMR (125 MHz, CDCl₃, δ _C) 166.2 (C=O), 166.0 (C=O), 165.5 (C=O), 165.1 (C=O), 164.9 (C=O), 155.1 (Ar), 150.2 (Ar), 133.6 (Ar), 133.5(7) (Ar), 133.5 (Ar), 129.9 (Ar), 129.8(5) (Ar), 129.8(3) (Ar), 129.8(1) (Ar), 129.6 (Ar), 129.1 (Ar), 129.0(7) (Ar), 129.0(3) (Ar), 128.8 (Ar), 128.5(8) (Ar), 128.5(5) (Ar), 128.5(4) (Ar), 128.5 (Ar), 128.3 (Ar), 118.3 (Ar), 114.6 (Ar), 105.3(6) (C-1), 105.3(1) (C-1), 105.1 (C-1), 84.7, 84.1, 82.8, 82.0, 81.4, 81.1, 80.1, 78.2, 77.6, 64.9 (C-5), 62.7 (C-5 \times 2), 55.6 (OCH₃); ESIMS *m/z* calcd for (M + Na) C₅₇H₅₂O₁₉ 1063.2990, found 1063.2995.

p-Methoxyphenyl 2,3-Di-O-benzoyl-5-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3-di-O-benzoyl-5-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (27). Compound **27** was prepared as a syrup (1.92 g, 94%) from thioglycoside **15** (1.77 g, 1.7 mmol), alcohol **26** (0.74 g, 0.71 mmol), 4 Å molecular sieves (0.65 g), *N*-iodosuccinimide (0.38 g, 1.7 mmol), and silver triflate (0.04 g, 0.15 mmol) in CH₂Cl₂ (55 mL) as described for **16**. The compound contained 5% of an inseparable impurity and was used directly in the next step. Data for **27**: *R_f* 0.33 (65:35 hexanes-EtOAc); [α]_D +17.8 (c 1.3, CHCl₃); ¹H NMR (600 MHz, CDCl₃, δ _H) 8.10–7.84 (m, 24 H, Ar), 7.73–7.66 (m, 8 H, Ar), 7.59–7.20 (m, 53 H, Ar), 7.01 (d, 2 H, *J* = 8.4 Hz, Ar), 6.75 (d, 2 H, *J* = 8.4 Hz, Ar), 5.76 (s, 1 H), 5.73 (d, 1 H, *J* = 4.5 Hz), 5.67 (d, 2 H, *J* = 5.8 Hz), 5.65 (d, 2 H, *J* = 4.7 Hz), 5.63–5.60 (m, 4 H), 5.58–5.54 (m, 4 H), 5.38–5.35 (m, 3 H), 5.33 (s, 1 H), 5.30 (s, 1 H), 4.63–4.58 (m, 3 H), 4.56–4.52 (m, 2 H), 4.50–4.44 (m, 3 H), 4.23–4.14 (m, 4 H), 4.06 (dd, 2 H, *J* = 11.5, 4.5 Hz), 4.02–3.94 (m, 4 H), 3.92–3.84 (m, 4 H), 3.82 (dd, 1 H, *J* = 11.2, 2.7 Hz), 3.73 (s, 3 H, OCH₃), 1.01 (s, 18 H, C(CH₃)₃ \times 2); ¹³C NMR (125 MHz, CDCl₃, δ _C) 165.5(7) (C=O), 165.5(6) (C=O \times 3), 165.4(8) (C=O), 165.4(5) (C=O \times 2), 165.1(6) (C=O), 165.1(5) (C=O), 165.1(4) (C=O), 165.1(1) (C=O), 165.0(5) (C=O), 165.0(3) (C=O), 155.0 (Ar), 150.3 (Ar), 135.6(6) (Ar), 135.6(4) (Ar), 133.4 (Ar), 133.3(4) (Ar), 133.3(0) (Ar), 133.2(8) (Ar), 133.2(6) (Ar), 133.2(2) (Ar), 133.1(4) (Ar), 133.1(1) (Ar), 133.0 (Ar), 132.9 (Ar), 130.0 (Ar), 129.9(8) (Ar), 129.9(4) (Ar), 129.9 (Ar), 129.8(3) (Ar), 129.8(2) (Ar), 129.7(6) (Ar), 129.7(3) (Ar), 129.6 (Ar), 129.3 (Ar), 129.2 (Ar), 129.1(6) (Ar), 129.1(3) (Ar), 129.0(8) (Ar), 129.0(3) (Ar), 128.4(8) (Ar), 128.4(5) (Ar), 128.4(2) (Ar), 128.3(6) (Ar), 128.3(4) (Ar), 128.2(4) (Ar), 128.2(2) (Ar), 128.1 (Ar), 127.6 (Ar), 118.4 (Ar), 114.5 (Ar), 106.1 (C-1), 106.0 (C-1), 105.9 (C-1), 105.8 (C-1 \times 2), 105.4 (C-1), 105.3 (C-1), 83.2, 83.1(9), 83.1(7), 83.0, 82.8, 82.6, 82.5, 82.1(7), 82.1(5), 82.1(2), 82.0, 81.4(8), 81.4(5), 80.5, 77.3, 77.2(4), 77.2(2), 76.8, 65.8 (C-5), 65.7(9) (C-5), 65.7(4) (C-5), 65.6(7) (C-5), 65.6(2) (C-5), 63.4 (C-5), 55.6 (OCH₃), 26.7 (C(CH₃)₃ \times 2), 19.2 (C(CH₃)₃ \times 2); ESIMS *m/z* calcd for (M + Na) C₁₆₅H₁₅₂O₄₃Si₂ 2901.9, found 2902.0.

p-Methoxyphenyl 2,3,5-Tri-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3,5-tri-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (28). Compound **28** was prepared as a

syrup (1.12 g, 92%) from thioglycoside **19** (1.77 g, 3.11 mmol), acceptor alcohol **24** (0.35 g, 0.97 mmol), 4 Å molecular sieves (0.5 g), *N*-iodosuccinimide (0.7 g, 3.11 mmol), and silver triflate (0.05 g, 0.19 mmol) in CH₂Cl₂ (40 mL) as described for **16**: *R_f* 0.28 (7:3 hexanes-EtOAc); [α]_D +29.5 (c 0.9, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ _H) 8.10–7.90 (m, 14 H, Ar), 7.63–7.32 (m, 14 H, Ar), 7.29–7.20 (m, 7 H, Ar), 7.06–7.0 (m, 2 H, Ar), 6.80–6.74 (m, 2 H, Ar), 5.78 (s, 1 H, H-1), 5.68 (s, 2 H), 5.62–5.58 (m, 2 H), 5.56 (s, 1 H), 5.53 (d, 1 H, *J* = 4.4 Hz), 5.39 (s, 1 H, H-1), 4.77 (dd, 1 H, *J* = 11.4, 3.2 Hz), 4.72 (dd, 1 H, *J* = 11.4, 3.7 Hz), 4.70–4.52 (m, 6 H), 4.12 (dd, 1 H, *J* = 11.5, 4.2 Hz, H-5), 3.93 (dd, 1 H, *J* = 11.5, 2.6 Hz, H-5), 3.74 (s, 3 H, OCH₃); ¹³C NMR (125 MHz, CDCl₃, δ _C) 166.1 (C=O), 166.0 (C=O), 165.6 (C=O), 165.5(8) (C=O), 165.5(3) (C=O), 165.1 (C=O), 155.1 (Ar), 150.2 (Ar), 133.5(2) (Ar), 133.5(0) (Ar), 133.4 (Ar), 133.3 (Ar), 132.9 (Ar), 129.9(7) (Ar), 129.9 (Ar), 129.8 (Ar), 129.7(8) (Ar), 129.7(6) (Ar), 129.7(2) (Ar), 129.6 (Ar), 129.5 (Ar), 129.0(6) (Ar), 129.0(5) (Ar), 129.0(2) (Ar), 128.9 (Ar), 128.5(2) (Ar), 128.5 (Ar), 128.3 (Ar), 128.2 (Ar), 118.4 (Ar), 114.5 (Ar), 106.1 (C-1), 105.7 (C-1), 105.2 (C-1), 82.9, 82.5, 81.9, 81.7, 81.6, 80.8, 77.7(3), 77.7(2), 65.7 (C-5), 63.7 (C-5), 63.6 (C-5), 55.6 (OCH₃); ESIMS *m/z* calcd for (M + Na) C₇₁H₆₀O₂₁ 1271.3522, found 1271.3524.

p-Methoxyphenyl 3,5-O-(Di-tert-butylsilyl)-2-O-levulinoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[3,5-O-(di-tert-butylsilyl)-2-O-levulinoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (31). Compound **31** was prepared as a syrup (2.44 g, 76%) from thioglycoside **30** (4.61 g, 9.31 mmol), acceptor alcohol **24** (1.05 g, 2.91 mmol), 4 Å molecular sieves (1.5 g), *N*-iodosuccinimide (2.09 g, 9.3 mmol), and silver triflate (168 mg, 0.65 mmol) in CH₂Cl₂ (135 mL) as described for **16**: *R_f* 0.31 (7:3 hexanes-EtOAc); [α]_D +68.0 (c 0.8, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ _H) 8.08–8.04 (m, 2 H, Ar), 7.60–7.55 (m, 1 H, Ar), 7.48–7.43 (m, 2 H, Ar), 7.07–7.04 (m, 2 H, Ar), 6.84–6.80 (m, 2 H, Ar), 5.67 (s, 1 H), 5.54 (d, 1 H, *J* = 2.2 Hz), 5.22–5.18 (m, 2 H), 5.09 (dd, 1 H, *J* = 6.7, 2.1 Hz), 4.97 (d, 1 H, *J* = 2.0 Hz), 4.46–4.42 (m, 1 H), 4.35–4.28 (m, 3 H), 4.11 (dd, 1 H, *J* = 9.6, 7.0 Hz), 4.07 (dd, 1 H, *J* = 9.6, 6.7 Hz), 3.98 (ddd, 2 H, *J* = 10.3, 10.1, 5.5 Hz), 3.94–3.87 (m, 3 H), 3.80 (dd, 1 H, *J* = 11.4, 3.5 Hz), 3.76 (s, 3 H, OCH₃), 2.79–2.74 (m, 2 H, levulinoyl CH₂), 2.73–2.54 (m, 6 H, levulinoyl CH₂ \times 3), 2.14 (s, 3 H, levulinoyl COCH₃), 2.11 (s, 3 H, levulinoyl COCH₃), 1.05 (s, 9 H, C(CH₃)₃), 1.03 (s, 9 H, C(CH₃)₃), 0.99 (s, 9 H, C(CH₃)₃), 0.90 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ _C) 206.2 (C=O), 206.0 (C=O), 171.8 (C=O), 171.6 (C=O), 165.4 (C=O), 154.9 (Ar), 150.4 (Ar), 133.3 (Ar), 129.8 (Ar), 129.4 (Ar), 128.5 (Ar), 118.4 (Ar), 114.4 (Ar), 106.2 (C-1), 106.0 (C-1), 105.2 (C-1), 83.0, 82.8, 82.6, 81.7, 81.4, 80.4, 79.9, 73.7, 73.4, 67.4 (C-5), 67.3 (C-5), 66.0 (C-5), 55.6 (OCH₃), 38.0 (levulinoyl CH₂), 37.8 (levulinoyl CH₂), 29.7 (levulinoyl CH₂), 29.6 (levulinoyl CH₂), 27.9 (levulinoyl CH₂), 27.8 (levulinoyl CH₂), 27.3(9) (C(CH₃)₃), 27.3(7) (C(CH₃)₃), 27.0 (C(CH₃)₃), 26.9 (C(CH₃)₃), 22.6(3) (C(CH₃)₃), 22.6(0) (C(CH₃)₃), 20.0(8) (C(CH₃)₃), 20.0(0) (C(CH₃)₃); ESIMS *m/z* calcd for (M + Na) C₅₅H₈₀O₁₉Si₂ 1123.4725, found 1123.4724.

p-Methoxyphenyl 3,5-O-(Di-tert-butylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 3)-[3,5-O-(di-tert-butylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (32). A solution of compound **31** (3.46 g, 3.14 mmol) and hydrazine monohydrate-acetic acid (1:2, v/v; 42 mL) in THF (150 mL) and CH₃OH (15 mL) was stirred for 1.5 h. The solvent was removed, and the resulting oil was diluted with EtOAc (200 mL). The solution was washed with a saturated aqueous NaHCO₃ solution (150 mL \times 2) and brine (100 mL), dried (Na₂SO₄), and concentrated. The crude residue was purified by column chromatography (4:1 hexanes-EtOAc) to afford **32** (2.71 g, 95%) as a white solid: *R_f* 0.57 (3:1 hexanes-EtOAc); [α]_D +72.8 (c 0.3, CH₂-Cl₂); ¹H NMR (600 MHz, CDCl₃, δ _H) 8.06–8.04 (m, 2 H, Ar), 7.62–7.58 (m, 1 H, Ar), 7.48–7.44 (m, 2 H, Ar), 7.06–7.03 (m, 2 H, Ar), 6.85–6.82 (m, 2 H, Ar), 5.76 (s, 1 H, H-1), 5.53 (d, 1 H, *J* = 2.0 Hz, H-2), 5.18 (d, 1 H, *J* = 3.3 Hz, H-1'), 4.98 (d, 1 H, *J* = 3.3 Hz, H-1''),

4.46–4.43 (m, 1 H, H-4), 4.31–4.26 (m, 4 H, H-3, H-2', H-5' × 2), 4.12–4.10 (m, 1 H, H-2''), 4.02–3.89 (m, 7 H, H-5, H-3', H-3'', H-5'' × 2, H-4', H-4''), 3.78 (dd, 1 H, $J = 11.4, 3.2$ Hz, H-5), 3.77 (s, 3 H, OCH₃), 2.78 (d, 1 H, $J = 4.1$ Hz, OH), 2.49 (d, 1 H, $J = 4.2$ Hz, OH), 1.07 (s, 9 H, C(CH₃)₃), 1.05 (s, 9 H, C(CH₃)₃), 1.01 (s, 9 H, C(CH₃)₃), 0.93 (s, 9 H, C(CH₃)₃); ¹³C NMR (100 MHz, CDCl₃, δ_c) 165.6 (C=O), 155.1 (Ar), 150.1 (Ar), 133.5 (Ar), 129.8 (Ar × 2), 129.3 (Ar), 128.5 (Ar × 2), 118.4 (Ar × 2), 114.5 (Ar × 2), 108.3 (C-1'), 108.2 (C-1''), 105.1 (C-1), 83.6 (C-2), 82.5 (C-3), 82.2 (C-4), 81.4 (C-2', C-2''), 81.3(9) (C-4'), 81.3(5) (C-4''), 73.9 (C-3'), 73.7 (C-3''), 67.4(6) (C-5'), 67.4 (C-5''), 66.5 (C-5), 55.6 (OCH₃), 27.4(3) (C(CH₃)₃), 27.4(1) (C(CH₃)₃), 27.1 (C(CH₃)₃), 27.0 (C(CH₃)₃), 22.6(4) (C(CH₃)₃), 22.6 (C(CH₃)₃), 20.1 (C(CH₃)₃), 20.0 (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₄₅H₆₈O₁₅Si₂ 927.3989, found 927.3992.

p-Methoxyphenyl 2-O-Benzyl-3,5-O-(di-tert-butylsilanediyl)- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-O-(di-tert-butylsilanediyl)- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2-O-benzyl-3,5-O-(di-tert-butylsilanediyl)- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-O-(di-tert-butylsilanediyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (34). To a mixture of **32** (0.24 g, 0.265 mmol), **33**⁴¹ (0.39 g, 0.8 mmol), and 4 Å molecular sieves (0.4 g) in CH₂Cl₂ (40 mL) was added NIS (0.18 g, 0.8 mmol) followed by AgOTf (20 mg, 0.08 mmol) at –30 °C. After being stirred for 30 min, the solution turned dark red, and Et₃N was added. The mixture was then diluted with CH₂Cl₂ (20 mL) and filtered through Celite. The filtrate was washed with a saturated aqueous Na₂S₂O₃ solution (40 mL), dried (Na₂SO₄), and concentrated to give a crude residue that was purified by column chromatography (10:1 hexanes–EtOAc) to afford **34** (0.3 g, 70%) as a white semisolid: R_f 0.29 (8:1 hexanes–EtOAc); $[\alpha]_D -12.1$ (c 0.7, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃, δ_H) 8.01–7.99 (m, 2 H, Ar), 7.58–7.55 (m, 1 H, Ar), 7.45–7.21 (m, 12 H, Ar), 7.02–6.98 (m, 2 H, Ar), 6.81–6.77 (m, 2 H, Ar), 5.71 (s, 1 H, H-1), 5.49 (d, 1 H, $J = 1.5$ Hz, H-2), 5.32 (d, 1 H, $J = 3.1$ Hz, H-1'), 5.20 (d, 1 H, $J = 4.8$ Hz, H-1''), 5.03 (d, 1 H, $J = 2.7$ Hz, H-1'''), 4.97 (d, 1 H, $J = 4.8$ Hz, H-1'''), 4.85 (d, 1 H, $J = 12.5$ Hz, PhCH₂), 4.81 (d, 1 H, $J = 12.5$ Hz, PhCH₂), 4.73 (d, 1 H, $J = 12.3$ Hz, PhCH₂), 4.70 (d, 1 H, $J = 12.3$ Hz, PhCH₂), 4.50 (dd, 1 H, $J = 9.2, 9.2$ Hz), 4.45–4.41 (m, 2 H), 4.32–4.24 (m, 5 H), 4.19 (dd, 1 H, $J = 7.6, 3.1$ Hz, H-2'), 4.12–4.03 (m, 3 H), 4.00–3.86 (m, 8 H), 3.79–3.70 (m, 3 H), 3.75 (s, 3 H, OCH₃), 3.63–3.58 (m, 1 H), 1.09 (s, 9 H, C(CH₃)₃), 1.07 (s, 9 H, C(CH₃)₃), 1.05 (s, 9 H, C(CH₃)₃), 1.03 (s, 9 H, C(CH₃)₃), 1.01 (s, 9 H, C(CH₃)₃), 1.00 (s, 9 H, C(CH₃)₃), 0.99 (s, 9 H, C(CH₃)₃), 0.92 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.3 (C=O), 155.0 (Ar), 150.2 (Ar), 138.0 (Ar), 137.7 (Ar), 133.4 (Ar), 129.8 (Ar × 2), 129.1 (Ar), 128.4 (Ar × 2), 128.3 (Ar × 2), 128.2 (Ar × 2), 128.1 (Ar × 2), 127.9 (Ar), 127.7(4) (Ar), 127.7(1) (Ar), 127.5 (Ar), 118.1 (Ar × 2), 114.5 (Ar × 2), 107.2 (C-1'), 106.5 (C-1''), 105.0 (C-1), 99.6 (C-1'''), 99.4 (C-1'''), 86.4, 85.8, 83.3, 82.4, 81.5, 80.5, 80.4, 80.1, 79.8, 78.1, 78.0, 74.3, 74.1(9), 74.1(7), 73.9, 71.8 (PhCH₂), 71.6 (PhCH₂), 68.8, 68.7, 67.5, 67.4, 66.4, 55.6 (OCH₃), 27.8 (C(CH₃)₃), 27.5(7) (C(CH₃)₃), 27.5(6) (C(CH₃)₃), 27.4(3) (C(CH₃)₃), 27.4(1) (C(CH₃)₃), 27.2 (C(CH₃)₃), 27.1 (C(CH₃)₃), 27.0 (C(CH₃)₃), 22.6 (C(CH₃)₃ × 2), 22.5(9) (C(CH₃)₃), 22.5(8) (C(CH₃)₃), 20.1(3) (C(CH₃)₃), 20.1(1) (C(CH₃)₃), 20.0(7) (C(CH₃)₃), 20.0(1) (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₈₅H₁₂₈O₂₃Si₄ 1651.7827, found 1651.7827.

p-Methoxyphenyl 3,5-O-(Di-tert-butylsilanediyl)- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-O-(di-tert-butylsilanediyl)- α -D-arabinofuranosyl-(1 \rightarrow 3)-[3,5-O-(di-tert-butylsilanediyl)- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-O-(di-tert-butylsilanediyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (35). To a solution of compound **34** (0.24 g, 0.15 mmol) in EtOAc (10 mL) was added 10% Pd/C (70 mg), and the reaction mixture was stirred vigorously under hydrogen (1 atm) for 7 h. The reaction mixture was diluted with EtOAc (5 mL) and filtered to remove the catalyst. The filtrate was concentrated to give a syrup that was purified by column chromatography (9:1 hexanes–EtOAc) to afford **35** (0.18 g, 84%) as a syrup: R_f 0.43 (4:1 hexanes–

EtOAc); $[\alpha]_D -9.7$ (c 0.7, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.10–8.06 (m, 2 H, Ar), 7.62–7.56 (m, 1 H, Ar), 7.48–7.42 (m, 2 H, Ar), 7.05–7.00 (m, 2 H, Ar), 6.86–6.82 (m, 2 H, Ar), 5.78 (s, 1 H, H-1), 5.52 (d, 1 H, $J = 2.4$ Hz), 5.25 (d, 1 H, $J = 4.7$ Hz, H-1''), 5.22 (d, 1 H, $J = 3.2$ Hz, H-1'), 5.05 (d, 1 H, $J = 5.0$ Hz, H-1'''), 5.01 (d, 1 H, $J = 2.8$ Hz, H-1''), 4.39 (ddd, 1 H, $J = 6.5, 3.9, 2.8$ Hz), 4.32 (dd, 1 H, $J = 9.1, 5.1$ Hz), 4.30–4.14 (m, 7 H), 4.09–3.84 (m, 12 H), 3.77 (s, 3 H, OCH₃), 3.78–3.72 (m, 2 H), 3.64 (ddd, 1 H, $J = 10.5, 8.9, 5.0$ Hz), 3.24 (d, 1 H, $J = 9.6$ Hz, OH), 2.49 (d, 1 H, $J = 9.7$ Hz, OH), 1.09 (s, 9 H, C(CH₃)₃), 1.06 (s, 9 H, C(CH₃)₃), 1.04 (s, 9 H, C(CH₃)₃), 1.02 (s, 9 H, C(CH₃)₃), 1.01 (s, 9 H, C(CH₃)₃), 0.99 (s, 18 H, C(CH₃)₃), 0.89 (s, 9 H, C(CH₃)₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 165.9 (C=O), 155.1 (Ar), 150.1 (Ar), 133.7(Ar), 129.9 (Ar), 129.0 (Ar), 128.5 (Ar), 118.1 (Ar), 114.6 (Ar), 107.3 (C-1'), 106.9 (C-1''), 104.8 (C-1), 100.5 (C-1'''), 99.8 (C-1'''), 87.5, 87.2, 84.3, 83.3, 81.6, 80.3, 80.0, 79.7, 79.7, 75.7, 75.5, 74.4, 74.0, 73.9(8), 73.9(2), 68.6 (C-5), 68.4 (C-5), 67.4 (C-5 × 2), 65.8 (C-5), 55.6 (OCH₃), 27.6 (C(CH₃)₃), 27.5 (C(CH₃)₃), 27.3(4) (C(CH₃)₃), 27.3(1) (C(CH₃)₃), 27.1(9) (C(CH₃)₃), 27.1(6) (C(CH₃)₃), 27.1(1) (C(CH₃)₃), 26.9 (C(CH₃)₃), 22.6(9) (C(CH₃)₃), 22.6(8) (C(CH₃)₃), 22.6(5) (C(CH₃)₃), 22.6(4) (C(CH₃)₃), 20.1(5) (C(CH₃)₃), 20.1(1) (C(CH₃)₃), 20.0 (C(CH₃)₃), 19.9 (C(CH₃)₃); ESIMS m/z calcd for (M + Na) C₇₁H₁₁₆O₂₃Si₄ 1471.6880, found 1471.6882.

p-Methoxyphenyl 2,3,5-Tri-O-benzoyl- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 3)-[2,3,5-tri-O-benzoyl- β -D-arabinofuranosyl-(1 \rightarrow 2)-3,5-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)]-2-O-benzoyl- α -D-arabinofuranoside (36). To a solution of compound **35** (0.19 g, 0.13 mmol) in THF–pyridine (2:1; 12 mL) at 0 °C was added HF–pyridine (0.4 mL), and the resulting mixture was stirred for 20 h. The reaction mixture was diluted with a solution of DMF–pyridine–EtOAc (15:5:5, 25 mL), and solid NaHCO₃ was added in portions under vigorous stirring until the solution became neutral (stirred for 1 h). The reaction mixture was filtered, and the solids were washed with DMF–pyridine–EtOAc (15:5:5, 15 mL). The combined organic phase was concentrated to give a syrup that was quickly filtered through a short silica gel column. The fractions containing the pentasaccharide (R_f 0.23, 3:1 CH₂Cl₂–CH₃OH) were concentrated to give a syrup that was dissolved in pyridine (15 mL), followed by the addition of benzoyl chloride (0.2 mL, 1.7 mmol) at 0 °C. The resulting reaction mixture was stirred for 12 h and diluted with CH₂Cl₂ (30 mL). Chilled water (20 mL) was then added, and after the reaction mixture was stirred for 15 min, the separated organic layer was washed successively with a saturated aqueous NaHCO₃ solution (20 mL × 2) and water (20 mL), dried (Na₂SO₄), and concentrated to give a syrup that was purified by column chromatography (7:3 hexanes–EtOAc) to afford **36** (0.18 g, 70% over two steps) as a syrup: R_f 0.24 (65:35 hexanes–EtOAc); $[\alpha]_D -30.5$ (c 0.8, CHCl₃); ¹H NMR (500 MHz, CDCl₃, δ_H) 8.12–7.80 (m, 22 H, Ar), 7.61–7.56 (m, 2 H, Ar), 7.50–7.13 (m, 31 H, Ar), 7.04–6.98 (m, 2 H, Ar), 6.80–6.75 (m, 2 H, Ar), 5.95 (dd, 2 H, $J = 11.7, 5.3$ Hz), 5.84 (d, 1 H, $J = 4.8$ Hz, H-1''), 5.74 (s, 1 H, H-1), 5.71 (d, 1 H, $J = 4.8$ Hz, H-1'''), 5.57 (dd, 1 H, $J = 6.4, 4.8$ Hz), 5.47–5.42 (m, 3 H, H-1''), 5.37–5.33 (m, 2 H), 5.11 (s, 1 H, H-1'), 4.82 (dd, 1 H, $J = 11.7, 4.6$ Hz), 4.79–4.70 (m, 2 H), 4.69–4.62 (m, 2 H), 4.55 (ddd, 1 H, $J = 7.5, 6.4, 4.8$ Hz), 4.52 (d, 1 H, $J = 1.6$ Hz), 4.50–4.32 (m, 7 H), 4.18 (ddd, 2 H, $J = 11.3, 6.7, 4.7$ Hz), 3.97 (dd, 1 H, $J = 11.9, 4.0$ Hz), 3.77 (dd, 1 H, $J = 11.8, 2.3$ Hz), 3.74 (s, 3 H, OCH₃); ¹³C NMR (125 MHz, CDCl₃, δ_c) 166.0 (C=O), 165.9(7) (C=O), 165.9(2) (C=O), 165.8(8) (C=O), 165.8(4) (C=O), 165.7(4) (C=O), 165.7(3) (C=O), 165.6 (C=O), 165.3 (C=O × 3), 155.0 (Ar), 150.3 (Ar), 133.5 (Ar), 133.4 (Ar), 133.3 (Ar), 133.2(9) (Ar), 133.2(2) (Ar), 133.1 (Ar), 132.8(6) (Ar), 132.8(3) (Ar), 132.8(1) (Ar), 129.8(9) (Ar), 129.8(7) (Ar), 129.8(5) (Ar), 129.8(1) (Ar), 129.7(8) (Ar), 129.7(3) (Ar), 129.7(1) (Ar), 129.6(9) (Ar), 129.6(6) (Ar), 129.6(2) (Ar), 129.2 (Ar), 129.1 (Ar), 129.0 (Ar), 128.9 (Ar), 128.7 (Ar), 128.4(9) (Ar), 128.4(7) (Ar), 128.4(5) (Ar), 128.4(1) (Ar), 128.3 (Ar), 128.1(9) (Ar), 128.1(7) (Ar), 118.3

77.1, 75.7, 75.6, 75.0(6), 75.0(1), 69.4, 67.7 (octyl OCH₂), 67.6 (C-5), 67.5 (C-5), 67.3 (C-5), 67.2 (C-5), 67.1 (C-5), 63.8(6) (C-5), 63.8(2) (C-5), 61.4(9), 61.4(7), 52.1 (octyl CH₂N₃), 29.4 (octyl CH₂), 29.0(8) (octyl CH₂), 29.0(3) (octyl CH₂), 28.8 (octyl CH₂), 26.7 (octyl CH₂), 25.9 (octyl CH₂); ESIMS *m/z* calcd for (M + Na) C₁₁₈H₁₉₃N₃O₈₉ 3100.1, found 3100.0.

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Supporting Information Available: Experimental details and characterization data for the preparation of monosaccharide building blocks **10–12**, **20–24**, and **30** and ¹H and ¹³C NMR spectra of all previously unreported compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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